

Monoamine oxidase inhibitors: A review concerning dietary tyramine and drug interactions

Au: P Ken Gillman

PsychoTropical Research, Bucasia, Qld

Orchid ID 0000-0001-8277-3397

Researchgate https://www.researchgate.net/profile/Ken_Gillman

<https://psychotropical.com>

Abstract

This comprehensive monograph surveys original data on the subject of both dietary tyramine and drug interactions relevant to **Monoamine Oxidase Inhibitors (MAOIs)**, about which there is much outdated, incorrect, and incomplete information in the medical literature and elsewhere. Many of the drug interactions previously supposed to be serious are either non-existent or non-serious.

Very few foods now contain problematically high tyramine levels, that is a result of great changes in international food production methods and hygiene regulations. Cheese is the only food that, in the past, has been associated with documented fatalities resulting from acute tyramine-induced hypertension. Nowadays most cheeses are quite safe, and even ‘matured’ cheeses are usually safe in healthy-sized portions. The variability of ‘pressor’ sensitivity to tyramine between individuals, and the unpredictable amount of tyramine content in a few foods, means a little knowledge and care are still required.

The few interactions between MAOIs and other drugs are now well understood and are quite straightforward to avoid and deal with. They are detailed and discussed here (by a recognised expert). MAOIs have no clinically relevant *pharmacokinetic* interactions. The only significant *pharmacodynamic* interaction, other than the pressor response to tyramine (‘cheese reaction’), is serotonin toxicity (ST) — aka serotonin syndrome. That is now well defined and is straightforward to avoid by not co-administering any drug with serotonin re-uptake inhibitor (SRI) potency. There are no *therapeutically* used drugs, other than drugs with SRI activity, that are capable of inducing serious ST with MAOIs. Anaesthesia is not difficult or contra-indicated if a patient is taking MAOIs.

MAOIs are safe and straightforward to use, quite contrary to current popular opinion. Previously held concerns about MAOIs are mostly ‘mythical’ and misleading: either they are of over-rated importance, or incorrect, or no longer relevant.

Key words

Monoamine oxidase inhibitors, hypertension, drug interactions, serotonin toxicity, decarboxylating enzymes, biogenic amines, washout intervals, tricyclic anti-depressants (TCAs), clomipramine, imipramine, tranylcypromine, phenelzine and isocarboxazid, isoniazid, narcotic analgesics, triptans, histamine, putrescine, cadaverine, tyramine, tryptamine, 2-phenylethylamine, spermine, spermidine, methylamine, trimethylamine, scombroidosis, hypertensive urgency, hypertensive

This version July 2019 Check web site for latest V of this monograph

<http://www.psychotropical.com/anti-depressants/maois>

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

emergency, sub-lingual nifedipine, subarachnoid haemorrhage, end-organ damage, anaesthesia, indirect sympathomimetic activity, adrenaline, noradrenaline, dietary tyramine, cheese reaction, L-DOPA, dopamine, chocolate, wine, beer, chianti, aged cheeses, cured meats, pepperoni, salami, sauerkraut, kimchee, soy sauce, miso, fish sauce, yeast-extract spreads, health supplements, Marmite, broad bean pods, fava beans

Abbreviations and synonyms

adrenaline (adrenalin or epinephrine), noradrenaline (noradrenalin or norepinephrine), serotonin (5-HT), serotonin/noradrenaline re-uptake inhibitor (SRI, NRI) or serotonin/ noradrenaline transporter (5HTT or SERT/NAT or NET), monoamine oxidase inhibitors (MAOIs), serotonin toxicity (ST), Biogenic amines (BAs), tricyclic anti-depressants (TCAs), indirect sympatho-mimetic activity (ISA), 3,4-methylenedioxy-methamphetamine (MDMA, ecstasy), reversible inhibitors of monoamine oxidase A (RIMAs).

Introduction and background

If a man is offered a fact which goes against his instincts, he will scrutinize it closely, and unless the evidence is overwhelming, he will refuse to believe it. If, on the other hand, he is offered something which affords a reason for acting in accordance with his instincts, he will accept it even on the slenderest evidence. The origin of myths is explained in this way.
Bertrand Russell, Proposed Roads to Freedom

This monograph covers diet (both food and drink) and drug interactions for those on MAOIs. It is intended to update, inform, and assist both medical and non-medical readers.

It is lengthy, not because the subject is difficult or complicated, but because exposing the myths surrounding MAOIs involves more than just contradiction. The subject of MAOIs is richly cloaked in myth and unfortunately a mythical assertion often repeated is more firmly established in the minds of uncritical thinkers than a truth stated but once. Indeed, White and Simpson stated clearly 40 years ago in a report about TCA/MAOI combinations, adopted as ACNP policy, (A report to the American College of Neuropsychopharmacology [1]): that, ‘the riskiness of the combination is largely superstition’.

Interactions between **monoamine oxidase inhibitors (MAOIs)** and other drugs are not a difficult problem, contrary to current popular wisdom. These interactions have been well understood for some time, but that knowledge has been painfully slow to percolate to clinical practice. I have published various scientific papers relevant to this topic and am an internationally recognised expert on **serotonin toxicity (ST)** aka ‘serotonin syndrome’, which is the most serious potentially fatal interaction (see especially [2-10]).

MAOI interactions are neither frequent nor difficult to deal with, contrary to the impression generated by many standard texts. Experience shows problems and risks with MAOIs are less common than with SSRIs like fluoxetine, which has multiple potentially problematic interactions and yet is still widely used [11]. Side effects are frequently less with tranylcypromine than with SSRIs, which is reflected by patients’ generally stated preferences concerning optimum treatments [12-14].

Check web site for latest V

2

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

Standard texts cover many issues within tight space constraints. They contain abbreviated discussion and information, often by authors who are not really expert in the subject, that causes confusion because such texts contradict the contents of detailed analyses, such as this monograph.

There is now a lot of quality data on tyramine in foods, and also on how much tyramine is likely to constitute a potentially serious problem [15]. Previous opinions and advice have been based on old and sometimes inaccurate data, e.g. [1].

This monograph surveys more original data on tyramine than any paper previously published. There are more than 200 new references, mostly recent, that have never been cited in the medical literature.

Some of this monograph has now (2019) been published as a peer-reviewed article in the Journal of Neural Transmission [9].

Biogenic amines (BAs), including tyramine, are heat stable: they are unaffected by cooking. Furthermore, decarboxylating enzymes are also heat-tolerant and may survive some cooking, allowing continued accumulation of BAs if cooked food is then poorly refrigerated. Storage of foods below 5°C is a crucial factor, and some domestic fridges fail to maintain temperatures of below 5°C. Fridge temperatures must be checked with an accurate thermometer.

Some common myths

Science must begin with myths, then progress to the criticism of myths.
Karl Popper

All the following statements are wrong:

- The diet is difficult
- People cannot have cheese or red wine
- MAOIs have many dangerous interactions with other drugs
- One cannot give an anaesthetic
- It is difficult to swap to and from other drugs
- One cannot combine them with tricyclic anti-depressants (TCAs)
- They have a lot of side effects
- MAOIs cause hypertension/should not be given to hypertensive patients
- Tyramine reactions are dangerous and need urgent treatment
- Patients should be given nifedipine sub-lingually
- Adrenaline cannot be used

Units of measurement and quantities

All concentrations are given as milligrams (mg) of tyramine per kilogram (kg) or litre (L). Most food labels are legally obliged to quote information as content per 100 grams (abbreviation 100 g). Other abbreviations like: G, gm, gms and grms, are used, but 'g' is generally considered the correct notation.

Those living in non-metric areas will find it helpful learn to work in metric units: it is confusing to use standard servings/standard drinks or oz./pints. Some scientific

Check web site for latest V

3

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

papers still use different units of measurement in the same sentence (a patient weighing 180 pounds took a dose of 150 mg of a drug). That is like being told that someone is one meter 32.9 inches tall. Such practices are ultimately dangerous to people's lives. Especially for those in the USA, see the US metric association information:

<http://lamar.colostate.edu/~hillger/common.html>

and see also

<http://www.newscientist.com/article/mg20827840.200-uks-chief-measurer-unite-the-world.html>

How much tyramine produces a risk of serious hypertension?

For a majority of those who already follow healthy eating amounts and patterns the low tyramine diet involves almost no changes at all.

Although a small percentage of people may get a significant, but not serious or 'risky', blood pressure elevation with only 10 mg of tyramine, a substantial proportion of people need to have closer to 50 mg (in a meal) to get a **blood pressure (BP)** increase meeting the definition of a 'hypertensive urgency' (i.e. systolic blood pressure [SBP] >180 mm Hg). BP levels in 'hypertensive urgency' range from 180–220 mm/Hg **and do not engender an acute risk of serious consequences** (e.g. sub-arachnoid haemorrhage).

For a detailed analysis of the evidence relating to tyramine 'dose' and blood pressure [see below](#), and Gillman [2, 9]. Nevertheless, it is prudent to keep in mind that human responses to drugs and drug interactions vary from one individual to another and that there will always be exceptions to generalisations about doses and responses: that is one reason it is wise to monitor sitting & standing BP before and during treatment. See this pdf for instructions about BP monitoring:

http://www.psychotropical.com/images/pdf-downloads/MAOI_treatment_BP_so.pdf

It is easy to work out how much tyramine is in 10 or 100 grams or milliliters of any of these foods. People should familiarize themselves with what 10 g and 100 g looks like, and what sensible food portion sizes are. Those who eat 1 kg beef steaks, or half a kilo of cheese etc. will need to adjust to avoid trouble (and to be healthy). Some people (those with a BMI of more than 26) may benefit by consulting a dietician for explanations and education about how to eat healthily.

BMI (body mass index) is weight in kg divided by height in meters squared. i.e. for an average man = $70 \text{ (kg)} / 1.7 \text{ (m)}^2$; or $70 / 2.89 = 24.22$ Also see website information like <http://www.win.niddk.nih.gov/publications/PDFs/justenough.pdf>

Healthy portion sizes of cheese are approximately what is safe tyramine-wise: i.e. 100 grams of cheese in a meal is an unhealthily large portion. A healthy portion is 25 grams. Few cheeses (even 'mature' cheeses) contain more than 25 mg of tyramine in 100 grams (25 mg in 100 g = 250 mg/kg). So, a 25 g portion contains only 6 mg of tyramine and that will not cause a significant blood pressure rise, even in tyramine-sensitive individuals. Matured cheeses contain between 2 and 3.5 g of salt per 100 g [16], or 20–35 g/kg. The recommended daily sodium intake has now been reduced by

some authorities to 2 g daily (this is equivalent to 5 g of table salt). Thus, 30 g of a typical cheese provides 1 g of salt.

Even if excessive tyramine is ingested and BP increase occurs, serious consequences are very unlikely. With present-day lower tyramine foods, it is difficult, if not impossible, to ingest the massive quantities of tyramine that led to patient deaths in decades gone by.

If excess tyramine is ingested it will usually mean nothing more than monitoring blood pressure for a 1–4 hours. In many mild ingestions, which is all one is likely to see nowadays, BP will peak at about one hour.

Hasty and alarmist treatment of high BP by inexperienced doctors produces a risk of doing harm. Current expert opinion strongly advocates that hypertensive urgencies should be treated conservatively [9]. The safest ‘out-of-hospital’ intervention is a sedative dose of a benzodiazepine, see below, ‘Hypertensive urgencies and emergencies due to tyramine, and the questionable relationship to subarachnoid haemorrhage’, for details.

Monitoring blood pressure while on MAOIs

It is quite simple to monitor blood pressure with an electronic BP monitoring device (upper-arm, or wrist cuff). All those on MAOIs should be schooled to keep a blood pressure record from the beginning of treatment (including a one-week pre-treatment base-line record). The decrease in blood pressure on standing is a good indication of whether MAOIs are having a sufficient effect [9]. There is further explanation on Psychotropical.com explaining blood pressure monitoring and MAOIs.

http://www.psychotropical.com/images/pdf-downloads/MAOI_treatment_BP_so.pdf

There are good reasons for blood pressure monitoring.

In my experience it is not possible to make logical and timely decisions about dose adjustment unless the BP is properly monitored. BP drop on standing (two measurements are essential) is the best measure of the effectiveness of a given dose and essential to optimal speed of adjustment to the final effective dose, whilst avoiding problems of excessive faintness, or even falling, from postural hypotension.

Although most people will only react to larger amounts of tyramine there is wide variation in the population and a minority will experience greater BP elevation with relatively smaller doses of tyramine. Therefore, monitoring the BP will soon reveal those who are in the tyramine-sensitive group and warn of the need for extra care about diet (or addition of a noradrenaline re-uptake inhibitor (NRI) [see below](#)).

Part 1

Introduction to dietary guide

‘Dis-moi ce que tu manges, je te dirai qui tu es’
(Tell me what you eat, and I will tell you what you are)
Anthelme Brillat-Savarin *Physiologie du Goût* 1825

The drugs discussed in this monograph belong to the group called **Mono-Amine Oxidase Inhibitors (MAOIs)**. The enzyme **Mono-Amine Oxidase (MAO)** has two

Check web site for latest V

5

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

sub-types, A and B. This information is most relevant for irreversible ‘non-selective’ MAO-AB inhibitors (the most common are **tranylcypromine, selegiline, phenelzine, and isocarboxazid**), and less important for various other types of MAOI.

Persons on these drugs may be advised to keep some means of identifying the fact that they are on MAOIs readily available. Similar steps as may be taken with insulin dependent diabetes and those suffering from epilepsy are appropriate; this is in case of accidents or emergencies. This might be a medical alert bracelet, and/or information in handbag or purse or wallet.

All treating medical practitioners should be informed if a patient is taking MAOIs, but most of them will not know what to do and will therefore offer inappropriate advice taken from out-of-date sources. Generally, advice on MAOIs should come from specialist psycho-pharmacologists, general psychiatrists usually have insufficient knowledge to manage MAOIs. Almost all the information on the Internet is significantly inaccurate, and even the information on sites of educational institutes is mostly out-dated and misleading [9].

The information provided here is authoritative; I have published multiple recent papers in prestigious scientific journals on the pharmacology of MAOIs and **tricyclic antidepressants (TCAs)** and their interactions and I have a great deal of first-hand practical experience, see especially references: [2-4, 6-9, 15, 17-20].

The mechanism of tyramine formation

Tyramine formation in foods requires the availability of the amino acid precursor tyrosine and the presence of micro-organisms with amino acid decarboxylase enzyme activity. If favourable conditions for their growth and decarboxylating activity exist then tyramine, and other **biogenic amines (BA)** like histamine, cadaverine and putrescine may accumulate in foods.

Tyrosine, but little or no tyramine, is present at up to 20 mg/kg in animal protein sources, but is generally lower in plants (see below for exceptions). That is why fresh properly cold-stored foods are always safe. Animal protein can accumulate tyramine if allowed to go ‘off’. Meat, fish etc. must be stored at a fridge temperature of less than 5°C. Meats that have been minced can be prone to bacterial contamination if unhygienically handled. Poorly handled mince that has been improperly refrigerated could accumulate significant tyramine quite quickly. That is why meat and fish processing must now take place at below 4°C by regulation in most countries. Few people in western society would now accept green rotten smelly meat but eating meat like that was common practice in times gone by, and still is in some places.

Histamine, putrescine, cadaverine, tyramine, tryptamine, 2-phenylethylamine, spermine and spermidine are the most important BAs in foods [21-25]; that is why smell (putrescine – putrid) is a helpful guide for what to avoid.

Measurement of biogenic amines

Much progress and refinement of measurement techniques of tyramine and biogenic amines in food has been made [26, 27]. Older estimations of tyramine concentrations may sometimes have been less accurate, especially because the isolation of amines from complex food matrices is not simple. Usually a derivatisation procedure needs to be applied to enable analysis by methods such as liquid chromatography (LC), or

gas chromatography (GC) with various detectors, including a mass spectrometer [21, 28-31]. Techniques are continuing to become better, faster, and less costly, so data are continuing to accumulate rapidly [21, 28-31]. Google scholar finds over 1000 references for 'biogenic amines food wine' between 2014 and 2015).

Toxicity of biogenic amines

Some BAs are toxic above a certain fairly low concentration. The amine most commonly implicated in toxicity in humans is histamine, which is responsible for the type of poisoning that occurs when spoiled fish is eaten. That is often called scombroidosis ([see below](#)). Recent reviews of the toxicity of amines give up-to-date information [21, 22, 32-34].

The symptoms of a hypertensive reaction

A reaction is a progressive increase of **blood pressure BP** over 30–60 minutes (faster for liquids taken on an empty stomach) and may manifest first as a forceful thumping heartbeat. The heart rate usually becomes slower [35-38], in response to the increase in BP. If **systolic blood pressure (SBP)** goes above around 180 mm Hg quite rapid onset of severe headache is usual (although headache is not a reliable indicator of high BP). Tightness in the chest, paleness (pallor) may occur.

The degree of increase in BP is proportional to the amount of tyramine ingested.

BP elevation starts soon after ingestion, usually around 30 – 60 minutes, and symptoms may occur soon after. Any symptoms, including headache, starting more than two hours after eating are less likely to be due to a hypertensive reaction as the duration of the reaction is usually not more than 1–2 hours.

An SBP of 180 mmHg or more, sustained over 3 measurements in 10 minutes or so, performed in a calm setting with an accurate sphygmomanometer is now referred to as a 'hypertensive urgency'. If 'end organ' dysfunction is present it is called a 'hypertensive emergency'. End organ dysfunction is uncommon unless **diastolic blood pressure (DBP)** is greater than 130 mmHg.

In hypertensive urgencies the treatment aim is to reduce BP slowly over 24–48 hrs. Since tyramine reactions are self-limiting over 2–4 hrs., even for moderately severe ones, it is clear they will very rarely require intervention.

Tyramine in foods and beverages

Myth: The diet is difficult. One cannot have cheese or red wine

General comments on diet and tyramine

'The pleasures of the table belong to all men and to all ages, and of all natures gifts remain the last, to console us for the passing of the rest.'
Anthelme Brillat-Savarin

This monograph reviews tyramine concentrations as indicated by a large body of food science research. Tyramine concentrations for ordinary foods depend on storage time and storage conditions. Modern food hygiene and handling practices and regulations in civilised countries mean that excess tyramine levels are unheard of in 'fresh' foods.

That leaves those foods that are deliberately produced using micro-organisms, that is the subject-matter of a major part of this monograph.

Minimising or avoiding the very few high tyramine foods and beverages that do exist is easy and necessary whilst taking MAOIs. Only a few foods can build up the degree of excess tyramine (hundreds of mg/kg) that can greatly elevate the BP. The result of any BP reaction is in proportion to the amount of tyramine that is consumed i.e. **BP elevation is a dose-related effect**: that is why it is permissible and safe to cautiously ‘test’ small quantities of some foods e.g. your favourite cheese.

This monograph cannot and does not deal individually with compound foods, e.g. pizza. Such foods can have various types of ingredients that may have widely different tyramine contents. The total tyramine content of such foods will depend on the individual ingredients, but a little common sense and calculation, from the information herein, will yield an estimate of the tyramine content.

Special starter cultures that have low levels of decarboxylating micro-organisms in them have been developed and are now used in almost all food production processes. They are used by most cheese-makers, partly because they minimise the formation of undesirable ‘off’ flavours. They also minimise the proliferation of undesirable contaminant organisms (cf. yoghurt, below) and thereby greatly lessen, or even prevent, tyramine formation. Worldwide, attention has focussed more on ‘food hygiene’ and the European Union have an extensive program of monitoring and research, e.g. see ‘Controlling Biogenic Amines in Traditional Food Fermentations’ [39], and under ‘Salami’ below.

http://cordis.europa.eu/result/rcn/86612_en.html

Tyramine only accumulates in significant quantities when tyrosine is converted to tyramine by decarboxylase enzymes possessed by some, but not all, micro-organisms (see e.g. [40]). The only foods that have enough tyramine in them to cause significant reactions are those that have been subjected to the action of these particular types of micro-organisms. However, modern food hygiene standards are such as to make that increasingly rare, because biogenic amines, including tyramine are monitored as part of food quality and hygiene audits [21, 41].

A potentially significant elevation BP can only occur if a relatively large amount of tyramine is eaten or drunk. For those on MAOIs most people (around 50% of the population) will need to ingest at least 25 mg of tyramine. A small proportion of people are more sensitive to tyramine and in such subjects 10 mg may be enough to cause a measurable or symptomatic BP elevation. Most foods with elevated tyramine (like matured cheeses) actually have no more than 250 mg/kg. Therefore, quantities of up to 100 g of such a cheese (and that is a large portion size), may be consumed without consequence by most people. Further discussion is in Finberg & Gillman [9, 15].

The earliest work on this subject remains instructive. Barry Blackwell described the cause-effect nature of the ‘cheese reaction’: The original papers by Blackwell [42-51], summarize very well most of the basic points that are in this monograph. For those who like to know more about history Blackwell has written about it recently on the web site of the recently founded ‘International Network for the History of Neuropsychopharmacology (INHIN)’ here:

Check web site for latest V

8

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

<http://www.inhn.org/controversies/barry-blackwell-adumbration-a-history-lesson.html>

'The wheel is come full circle, I am here'
Edmund, King Lear

Barry Blackwell has expressed his appreciation about this monograph and still feels that the chemist, Rowe, who made the key observation (his wife was taking 'Parnate'), has not had sufficient recognition. Although Barry described Rowe's role in the story in a subsequent paper, his name was not in the initial publication, and Barry told me he still regrets that. It is therefore satisfying to be able to 'close the circle' by refreshing our memories of the history of this subject in this review.

Seminal early research on the tyramine content of cheeses was done by Kosikowski, e.g. [52]. It is interesting to note that the series of papers he authored in the 1950s have never been cited in the medical literature, except by Blackwell [47]*. So, his efforts also have had insufficient recognition.

*derived from Google scholar 'cited by' links

Blackwell noted that almost all cases of the 'cheese reaction' then reported (1965) implicated cheddar cheese, some of which had been assayed as having around 3,500 mg/kg of tyramine [53], which greatly exceeds (by about two orders of magnitude) the values generally found in any assays of similar cheeses in the current era.

The explanation for the absence of data about tyramine in the medical literature is that medical writers have only searched for papers using the medical literature databases (i.e. 'PubMed' etc.). However, such databases do not include many of the food science related research journals, and it is in such journals that the data actually reside. A recent example of only citing papers from the medical literature databases is exemplified by the citations in Flockhart's 2012 review [54] and many others [55-58], which although presented as 'updates', has no original material about tyramine after 1996, and no data at all from the food science literature. Not much of an update.

Most of the references herein come from the food science literature, so a majority of them will not be located by a 'PubMed' search.

Only very rarely encountered foods will now have high tyramine concentrations, such as 1,000 mg/kg, or greater.

Dairy products, cheeses etc.

Cheese: 'Milks' leap towards immortality'
Clifton Fadiman

Most cheeses now have low tyramine levels (< 10 mg/kg), whether they are hard, semi-hard, acid-curd or soft [29, 31, 59-62].

It is likely that the unusually high concentrations of 1,000–3,000 mg/kg or more reported occasionally in older samples will no longer occur because food regulations have driven widespread reductions of tyramine levels, especially through the use of starter cultures [21, 62].

Matured and 'artisanal' cheeses can sometimes develop high concentrations of tyramine (~1,000 mg/kg), although many are surprisingly low. 'Matured' usually means aged for more than 3 months (typically 6 months or more), rather than just a few weeks.

Check web site for latest V

9

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

Contrary to what one might assume from the (lack of) data in the medical literature there have been thousands of tyramine estimations performed from cheeses all over the world: a selection of studies with extensive and varied sampling is given here to illustrate this.

Almost all commercial lower priced ‘processed’ and ‘supermarket’ cheeses are low in tyramine (always <200 mg/kg, usually in the range of 0 – 50 mg/kg) because ‘supermarket’ type outlets require large quantities of produce (i.e. industrial-scale, not artisan), and low prices do not pay for long warehouse ageing (i.e. more than 3 months).

Bunkova et al. recently reviewed the widely marketed Edam-style cheese [60]. As they point out:

“Optimum ripening time of these products is 6–10 weeks, usually at a temperature of 10-14 °C. However, nowadays, young cheeses (2-4 weeks old) are delivered to retail by many producers for economic reasons.”

They studied tyramine levels during maturation and storage and noted particularly that “In all ripening/storage regimes tested, the highest content of tyramine, putrescine and cadaverine was found in the edge [rind]. On the other hand, the lowest content was detected in the cheese core.”

They found that tyramine levels increased in approximately linear fashion over time, being 60 mg/kg at 60 days and reaching a maximum of 120 mg/kg at 100 days in the outer layer (rind) and in the core only 70 mg/kg.

Processed cheese

Processed cheese generally has low levels of tyramine. Ibrahim et al. analysed 45 samples of processed cheese made from a variety of types and found the mean was ~ 200 mg/kg for cheddar styles, and 100 mg/kg for Gouda styles, however, there were a small number of samples that were rather higher, the maximum being 800 mg/kg. [63]. Note, these were shelf samples from Egyptian retail outlets, who knows how long they had been on the shelf? Or at what temperature?

Classic matured (hard, semi-hard) cheeses

French

Francophiles may be surprised to be reminded that there are relatively few French hard cheeses and even fewer that are available outside France, examples are: Cantal, Comté, Emmental (generally produced industrially) and Mimolette (Edam-like).

Comté AOC: Mayer: tyramine 0 mg/kg [31]. Comté is essentially the same as Swiss Gruyere, but still mostly in the hands of small producers, whereas Swiss Gruyere is almost entirely large-scale co-operatives. One would therefore predict the Swiss types would be even lower in tyramine.

Cantal: Mayer: 0 mg/kg [31].

Italian

Parmigiano Reggiano: aged 24 and 30 months, tyramine 20 – 150 mg/kg [61], but Mayer [31] found levels < 10 mg/kg in the 6 samples he tested*

*Table 2; note the blank spaces in this table denote ‘undetectable’ as confirmed with Mayer (personnel communication).

Grano Padana (12 & 22 months old) all samples tyramine < 130 mg/kg. Mayer [31] found undetectable levels.

The Spizzirri paper included a wide range of cheeses (mostly Italian), Grana Padano, Pecorino, Provolone, Ripened goat cheese, Emmentaler, Taleggio, Bel Paese and more, none of which had more than 200 mg/kg of tyramine.

Italian pecorino [64]. This paper reviews tyramine levels in a wide variety of pecorino cheeses made from all the different significant producing regions of Italy. Some of them are very ‘artisan’ type cheeses and there is great variation. Many have quite low levels in the region of 100–200 mg/kg, but one particular example, Pecorino Del Parco Di Migliarino San Rossore, exceeds 1000 mg/kg.

British

Cheddar: young cheddar (4 weeks) all tyramine < 50 mg/kg, at 36 weeks maturation all samples < 160 mg/kg [65], and only 6 mg/kg [66] and Mayer [31] found levels of tyramine 0 mg/kg.*

*Note the great difference from the old assays, one or two orders of magnitude, < 50 mg/kg vs. the old value of around 3,500 mg/kg of tyramine [53, 67].

Dutch

Gouda is a very widely copied cheese style which when young is semi-soft and hardens with age. Real aged Dutch Gouda is called “Oude kaas (10–12 months old), Overjarige kaas (18 months old). Tyramine levels will vary with age, younger ones seem to have very low levels, older ones 100 – 250 mg/kg [31].

Dutch-type semi-hard cheeses mostly tyramine < 50 mg/kg, max 250 [68, 69].

Swiss

Gruyere: tyramine < 100 mg/kg [61]

Emmental: tyramine 0–68 mg/kg [31] and Spizzirri [61] 16 mg/kg.

Other cheeses (non-hard)

Brie and camembert styles (un-washed rind)

Normally these cheese styles (mould-ripened soft cheeses, fluffy ‘cotton-wool’ like surface) are only matured for 4 weeks before release, so low tyramine levels are expected. Tyramine concentrations are less now than in the past because of starter cultures and better storage (see below for older results). Thus, it is no surprise to find that the latest estimations using modern assay techniques give very low tyramine levels of < 10 mg/kg.

Mayer et al. looked at examples from Austria, Denmark and France and found negligible tyramine levels (maximum of 5 mg/kg) in 5 different types of un-washed rind soft cheeses [31]. Likewise, Bonczar [66] found only 6 mg/kg in three samples.

Some older papers have reported much higher levels which may be explained by poor production and storage and/or faulty assays; Horwitz [70, 71] found tyramine ~100 mg/kg. Other older papers found undetectable levels [52].

Colonna (1970s): Camembert (French), 20 samples, most had low levels of tyramine ~100 mg/kg, but showing large variation up to 1800 mg/kg [72].

Check web site for latest V

11

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

De Vuyst: Brie; tyramine 0 – 400 mg/kg, camembert very low, maximum 20 mg/kg [67].

Voight: Brie tyramine 0 – 260 mg/kg [73].

Washed-rind cheeses

Washed rind cheeses (Epoisses is the classic) encourage mould rather than bacteria and thus have low tyramine levels like Brie and Camembert Styles, Samkova [74] tested 30 samples and all had < 2 mg/kg. Coton found 30 mg/kg [75].

Smear ripened

Cheeses made in this style are not intended for long storage and a bacterium, often *Brevibacterium*, is smeared onto the rind of the cheese. These tend to have slightly higher tyramine levels, of 45 samples tested average levels were 150 mg/kg (max 500 mg/kg).

Others

Acid-curd cheeses. Some are coagulated (curdled) using rennet, but some undergo curdling by bacterial lactic acid fermentation, and these might be expected to contain a little tyramine. See below under ‘Austrian’ [59].

Feta style: generally low tyramine but ‘older’ examples creep up a bit to 250 mg/kg at 120 days of age [76].

Austrian

An extensive analysis in 2013 of 47 different Austrian cheeses, particularly ripened acid-curd cheeses, is detailed in Fiechter et al. [59]. Most have low tyramine levels of < 100 mg/kg, only 18/47 samples were > 100 mg/kg). The median concentration for tyramine was 30 mg/kg. One sample of aged acid curd (Ennstaler Steirerkäse with crumble texture) was the highest was at nearly 2,000 mg/kg).

Roquefort and Roquefort styles

Roquefort and Roquefort style ‘traditional’ cheeses (all made with *Penicillium roqueforti*), these include: Fourme d’Ambert, Bleu de Bresse, Gorgonzola, Stilton, Cabrales, Gamonedo and the ‘industrially-produced’ types Danish Blue, Bleu d’Auvergne, Edelplizkäse, Mycella.

Roquefort 4 samples: tyramine 0 mg/kg [31].

Blue cheese, Czech [77, 78]: the mean and median being tyramine 380 mg/kg and 289 mg/kg, respectively) and, different cheeses (vats) varied widely, from 10 mg/kg, to 875 mg/kg.

Non-matured cheeses, yogurt, milk

Cheese spreads

These occupy an in-between position in that it depends on what they are made from: some higher quality cheese spreads are made from proper vintage cheeses, a few of which may be relatively high in tyramine. As an example, ‘Parmareggio’ cheese spread clocked in at tyramine 40 mg/kg, not high, but significant if one was to eat a whole

tub of it [61]. On the other hand, most spreads are like commercial cream cheeses and contain no tyramine.

Unripened cheese styles

Fresh non-matured, i.e. unripened/unaged, cheese styles, and yoghurt, are always safe because milk itself has no tyramine, e.g. curd styles, *fromage frais*, mascarpone, cream, ricotta, mozzarella, cottage cheeses, bocconcini.

Spizzirri et al. assayed multiple samples, tyramine: 0 mg/kg [61].

Unripened cheeses: 10 samples [79] tyramine < 0.5 mg/kg.

Goats cheese [80], unripened 'frais' styles, usually tyramine < 5 mg/kg, many 0 mg/kg [61].

Aged goats' cheeses: usually low tyramine < 10 mg/kg [61], but some may be higher, e.g. 70 mg/kg [80].

Milk and yogurt

In France, the regulations are strict. To be called yoghurt milk must be fermented by *Lactobacillus bulgaricus* and *Streptococcus thermophilus* (no decarboxylase activity, so no tyramine), via starter cultures. Bacteria have to be at least at 10,000,000 CFU/g till the end of shelf-life. That means it is virtually impossible for tyramine producing bacteria to gain a footing: so, yoghurt has no tyramine. Novella-Rodriguez, 5 samples, no tyramine [79, 81].

Cho, Korea, Yoghurt, 8 samples, max tyramine of 4 mg/kg [82].

But, be warned, if you are holidaying in the Himalayas, watch out for Tibetan traditional fermented yak milk which may have tyramine 900 mg/L [83].

Fermented vegetables/cereals (Inc. sauces)

Fermented cereals: Background history

A little caution is appropriate regarding sourdough bread because it can accumulate tyramine to a level of several hundred mg/kg, as can other similarly prepared foodstuffs widespread in other countries (see below).

Almost from the dawn of agricultural practice humans have learnt to increase the palatability and digestibility of legumes (see Soya) and cereals through the use of fermentation, both with yeasts and bacteria. Yeast fermentation does not give rise to tyramine, but bacterial fermentation can do, depending on the types of bacteria and their decarboxylating activity.

The United Nations food and agriculture organisation (FAO) classify cereal-related products 1) on the basis of raw cereal ingredients:

- a) wheat-based foods e.g. bouza, kishk
- b) rice-based foods e.g. busa
- c) maize-based foods e.g. ogi, bread, kenkey
- d) millet-based foods e.g. kunuzaki
- e) sorghum-based foods e.g. pito, ogi, bogobe, kisra, burukutu, kisra, injera
- f) barley-based foods e.g. beer

2) on the basis of texture:

- a) liquid (gruel) e.g. ogi, mahewu, burukutu, pito, uji
- b) solid (dough) and dumplings e.g. kenkey, agidi
- c) dry (bread) e.g. kiswa, injera

There are many dozens of local names for such preparations, for details consult culinary works, Wikipedia, google etc.

Sourdough bread

In the modern world the most prominent cereal-related solid-food vestige of these ancient fermentation practices is sourdough bread. This differs from normal bread because it utilises bacterial activity in the starter culture for making the dough. Just as with all other fermentation techniques this will not produce significant levels of tyramine if the usual modern standardised starter cultures (with minimal decarboxylase activity) are used, as is now generally the case with commercial production. However, Artisan producers may well utilise cultures with greater decarboxylase activity. Therefore, their products may sometimes contain significant levels of tyramine.

Recent research indicates what would be expected by anybody who has understood the contents of this monograph. Preparations made with standardised starter cultures are generally low in tyramine but there are some exceptions, usually home-made and locally made Artisan-type produce. Rizzello found tyramine levels of around 700 mg/kg in sourdough fermented wheat germ [84].

Özdestan has investigated various similar Turkish foods and found lowish tyramine levels [85, 86]: like kumru (ten samples of from different manufacturers in Turkey) < 5mg/kg, shalgam (20 samples) < 50 mg/kg, and tarhana (20 samples) 50 – 100 mg/kg.

Marmite, Bovril, Promite, Vegemite etc.

It is likely that changes in the way these products are prepared in recent years have lowered the tyramine content; but there are not many measurements to rely on.

Marmite is made from the residual brewer's yeast and the first production facility was near the Bass beer brewery in Burton on Trent: production started in 1902. It had/has relatively high amounts of biogenic amines ~320 mg/kg of tyramine [87] and 650 mg/kg of tyramine [88]. Both those are rather less than Blackwell's original estimate [48, 49, 51] of around 1,500 mg/kg, which may represent a change in production technique, or inaccuracies in measurement. One would need to take 30 ml to get 10 mg tyramine, which is more than is usually consumed.

Marmite-like spreads are somewhat similar to soy sauce and 'miso' which also involve 'fermentation' of brews containing non-animal proteins. They are usually used in small amounts, which can be safely eaten. A teaspoon (5 ml) of 'Marmite' would have only $5/1000 \times 300$ mg of tyramine, i.e. only a couple of milligrams.

Soy bean products

All *fermented* soy bean products like sauce and paste are prone to have significant tyramine levels.

For a list of fermented soy bean products see Wikipedia
https://en.wikipedia.org/wiki/List_of_fermented_soy_products

Non-fermented products like (most) tofu have no tyramine [89].

Soy sauce, natto, miso and sufu etc.

Soy sauce is made from steamed soybeans, roast wheat, and Koji fungus, the moromi mash may then ferment for as much as 2 years after which it is filtered and pasteurised. Soya beans have no tyramine; it is produced slowly during the fermentation reaching typical concentrations of ~150 mg per kg (litre) after many months. Miso is similar. The story with these products is an echo of the fermented cereal picture, just with beans as opposed to grains, so levels may vary [90-92].

Japanese soy sauce: Maximum tyramine 940 mg/L (i.e. approx. 1 mg/ml). Most samples measured have ranged between 10-200 mg/L [93]. Maximum tyramine concentrations in the past may have been as high as 1000 mg/L, so 25 ml of that would have contained 25 mg of tyramine.

Most supermarket Soy sauces actually have tyramine levels around 100 mg/L.

Yongmeia [94], 40 samples of Chinese soy, mostly tyramine less than 200 mg/L (20 of the 40 were < 100 mg/kg). The total content for the five biogenic amines in these samples was 497 mg/L with a range from 41.7 to 1357 mg/L. The concentrations for each of the five amines were: tyramine 0–673 mg/L, histamine 0–592 mg/L, cadaverine 0–550 mg/L, spermidine 0–486 mg/L and spermine' 0–145 mg/L.

Stute [95], 23 samples soy, all low tyramine < 200, except one clocked a staggering 6,000 mg/kg (dead rat in the vat? or just a typo for 600?).

Miso, 5 samples tyramine ~20 mg/L [82], and Kung 40 samples: all < 50 mg/L [96].

Other soy derived products like miso soup and sufu [82, 97] generally have similar concentrations. Miso, 5 samples tyramine < 25 mg/kg [82], and soy sauce tyramine < 50 mg/kg [82]. Sufu from Taiwan [98], and Miso 40 samples tyramine all < 10 mg/kg [96] but some rather higher [99].

'Natto' is another fermented soya bean preparation that sometimes achieves high tyramine levels, although < 100 mg/kg is typical [92, 100].

Soybean pastes ('Doenjang' etc.), of 23 samples most had undetectable levels, but a couple were > 1,000 mg/kg [101].

Fermented sauces: Animal

Fish sauces

In classical Roman cooking fish sauce was called garum or liquamen. They are ubiquitous now, but have long been deeply rooted in Far Eastern cuisine. Seafood, often anchovy, is allowed to ferment ~140–200 days. Names: Nuoc-Mam (Vietnam), Nam-Pla (Thailand), Budu (Malaysia), or Patis (Philippines) ketjap-ikan (Indonesia), ngapi (Burma), ishiru or shottsuru (Japan), colombo-cure (India Pakistan), yeosu (China), aekjeot (Korea). For more see Wikipedia, and for recent reviews refs [23, 82, 95]. NB Cho is in Korean, but the tables of values are readable.

They will, like everything, vary a bit with producer and hygiene quality, but seem usually to be OK, 200–500 mg/kg (bearing in mind it is, like soy sauce, a condiment, so if used in modest amounts (no more than ~ 20 grams) will be safe [102].

Korean fermented fish products tyramine < 50 mg/kg [82], liquid fish sauce made from a variety of things, scallop, squid etc. tyramine average 350, max (anchovy) 600 mg/kg [82].

Stute [95], 45 commercial fish sauces from the Far East, most < 200 mg/kg, maximum 588 mg/kg for tyramine.

Worcestershire sauce is fermented and contains anchovies (at least the original ‘Lea & Perrins’ version). There are many different producers of such sauces called ‘Worcestershire’ or ‘Worcester’ and there is no data on their tyramine content, but it is reasonable to assume it will be variable and similar to other fish sauces, probably lower. If used in condiment quantities, it is unlikely to add a significant tyramine load to a meal.

Meat and fish products

Fresh and frozen meat and meat products are safe, but if they are not fresh, i.e. if they have been subject to decomposition by micro-organisms, then they could be risky. Fresh liver has no tyramine [103], but if stored badly or past its ‘use by’ date when purchased, and then kept in a domestic fridge that is not cold enough, may become risky [104, 105]. The Hedberg paper [104] is a great illustration of good observation and investigation.

Ordinary commercial beef is not usually aged, and concentrations of tyramine are likely to be < 10 mg/kg. Galgano, 7 mg/kg after 8 days at +4°C [106].

Similarly, liver patés (and similar meat or fish pastes) are safe if freshly made and properly refrigerated (i.e. *below* 4°C), especially because such foods are normally consumed in small portions. No specific modern data is available as yet, but the lessons enumerated herein tell us what is likely. Liver [107] has no tyramine, but once processed and contaminated with bugs it is an ideal culture medium, so any laxity in hygienic preparation, storage time and temperature will result in a steady increase in tyramine. Concentrations of tyramine 100 – 500 mg/kg are likely in contaminated and badly stored product after a week or two.

Meat, fresh

General

Fresh meats contain no significant amounts of tyramine, for a review of amines in meat (and vegetables) see especially Kalac [108-110]. Also, for discussion see [111-113].

Stored *chilled* meats are safe (i.e. < 10 mg/kg) [106, 109, 114]. Beef: stored at –18°C for 178 days, tyramine max tyramine <4 mg/kg [114, 115].

Poultry

Chicken: refrigerated for 20 days at a temperature of +4±1°C in a domestic refrigerator. Tyramine level at one day, 3 mg/kg, 20 days, 15 mg/kg [116-119]. Moreira found well stored product < 5 mg/kg.

Check web site for latest V

16

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

Poultry: insignificant levels [120-124].

Duck: tyramine 0 [123].

Minced and ground beef

Minced and ground beef and ‘hamburgers’ are potentially problematic because any contaminant bacteria are mixed into a medium (mince) with a large surface area, which may then be sub-optimally stored. It is therefore reassuring to find assays have found negligible levels of tyramine < 3 mg/kg [125]. It might also be observed that, in North America alone, they consume millions of burgers per year and there are no reports of tyramine reactions associated with burgers.

Beef

Beef (stored above 0°C) can have significant tyramine concentrations: stored at +4°C for 21 days, 60 mg/kg, and after 36 days at +4°C 120 mg/kg [112]. Such meat is usually only available in the restaurant trade (at a high price!) but could contribute to excessive tyramine intake as part of a gourmet meal. However, there are no reports of reactions with beef in 50 years (cf. liver, a couple of reports of reactions in 50 years).

Pork

Pork and fresh pork products, not surprisingly, have no tyramine [121-123].

Offal

Fresh offal contains no tyramine. Kidneys, liver, duck giblets etc [107, 121, 123, 126, 127], all had no tyramine.

Sausages, pâté, meat pastes

These have minimal tyramine unless poorly prepared or stored [128-130].

Meats, preserved

Dry-cured meats

As with all *dry cured* meat products (as opposed to fermented ones) only low concentrations of tyramine are expected, Lorenzo found < 5 mg/kg [131], which agrees with [132]. So ‘Parma ham’, pastirma, jamon, prosciutto, coppa etc. will all be safe.

Fermented sausages

Concentrations of tyramine depend, as would be predicted, on the hygienic quality of the meat used and the strains of bacteria involved. Those produced with frozen meat (low temperature processing) usually have maximum concentrations of about 100 mg/kg. The improved starter cultures, now widely used, show a lack of, or much diminished, amino acid decarboxylase activity which results in lower concentrations of BAs [40, 133-137].

In their 2003 paper, ‘Biogenic amines in dry fermented sausages: a review’ Suzzi reviewed 20 studies from all over Europe [138] and found tyramine was usually below 200 mg/kg, very few samples were higher [132]. Suzzi ‘*In the several reports concerning the Spanish dry fermented sausages Chorizo, Fuet, Sobrasada and Salsichon, tyramine was generally*

detected at the higher concentration (exceeding 600 mg/kg in some sausages with mean values of about 200 mg/kg).'

In Spanish fermented sausages Chorizo, Fuet, Sobrasada and Salsichon tyramine was detected at up to 600 mg/kg in some sausages, with mean values of about 200 mg/kg [139].

French sausages, both artisanal and industrial, had tyramine maxima of 270 mg/kg [138, 140].

Papavergou [141]: 50 samples of dry fermented sausages sold in Greece, mean 100, max 500 mg/kg.

Hygiene and low temperature processing continue to improve steadily, more recent surveys find generally lower concentrations [135, 142, 143].

Latorre-Moratalla et al. is a good recent review: they found an average of 150 mg/kg, max < 200 mg/kg. The study received financial support from the European community project: 'Assessment and improvement of safety of traditional dry sausages from producers to consumers' (QLK1 CT-2002-02240, Website: www.clermont.inra.fr/tradisausage/). It is a good example of the efforts being made to monitor and improve hygiene standards.

Preparations of stock cubes, powders, bouillon, etc.

These are not prepared by fermentation but are flavoured extracts and reductions, therefore they are most unlikely to be high in tyramine. Populin tested broths (homemade or canned products from the market), soups (ready-to-eat soups, condensed soups and creams), soup bases (bouillon cubes, pastes and granulated powders), sauces and salad dressings from the European and US markets [87]. They found none exceeded tyramine 10 mg/kg.

Fish

Fresh fish

Levels of both tyramine and histamine may be increased in poorly refrigerated produce. However, with fish spoilage it is notable that histamine can be greatly elevated without significant elevation of tyramine [144]. Many regulations limit histamine, to between 50 (USA) and 200 mg/kg (EU). Histamine itself causes Scombroidosis [24, 145], [see below](#). Freshness and low-temperature handling is everything, and quality control and screening of imported produce continues to be a powerful force for improving hygiene and handling world-wide [122, 146].

A recent review confirmed low tyramine levels in properly handled raw and processed seafood [145].

Fresh fish usually has 2 – 5 mg/kg tyramine [147]. Whole and filleted trout kept on ice for up to 18 days, max at 18 days was 7 mg/kg [148, 149]. Frozen fish 1 mg/kg [150].

Herring, fresh, stored on ice (i.e. ~ 2°C) < 5 mg/kg [151, 152]. Storage conditions varied a little, but histamine reached 400 mg/kg whilst tyramine was low. After reaching a maximum of 100 mg/kg after seven days, tyramine then decreased on storage to 15 mg/kg at 15 days.

Chilled fresh and frozen or thawed salmon [147, 153] had a max of 40 mg/kg at end of shelf life.

Concerning histamine in fish, see [154].

Cured fish

Various types of fish (especially salmon) are ‘cooked’ using food acids (see also ‘pickling’). The most widely known dish using this technique is Gravlax, gravad lax (and various other spellings and derivations) which originated in the Scandinavian countries and has been adopted in America, especially in Jewish culture where the name has transmuted to ‘lox’. The data elsewhere in this monograph allow confidence that fresh hygienically prepared fish done in this manner would be expected to be completely safe. However, as with vegetables, deliberately fermented, or matured, product may develop significant tyramine concentrations.

Smoked fish

Smoked salmon [155] dry-salted, traditional smoking, sliced, vacuum-packed stored nine days at 4°C and 19 at 8°C contained no tyramine.

Cold smoked salmon tyramine < 20 mg/kg [156].

Dried Fish

Dried salted Tuna roe, tyramine was 90 mg/kg [157].

Canned fish

Some canned samples reach tyramine 10 mg/kg, but that seems rare [158]. Max 70 mg/kg [159]. Histamine (some were > FDA limit of 50 mg/kg), one was 1,000 mg/kg of histamine*, see [160, 161].

*That level of histamine would likely precipitate ‘scombroidosis’ in someone on phenelzine.

Pickled fish

Pickled herring does not involve a fermentation process and such products are safe providing they are hygienically prepared from fresh fish. Modern food auditing processes controlling the hygiene of processing plants, and low temperature processing, suggests that all commercially available supplies are likely to be of good quality and therefore safe. As with vegetables (cf. sauekraut), product that has undergone a fermentation process is different, and can contain significant concentrations of tyramine, like the Strömming (herring) in Baltic countries, which is fermented. So, the Norwegians have their rakfisk (fermented fish), and the Swedish fermented herrings (Surströmming), Icelanders fermented shark (Hákarl or kæstur hákarl), and perhaps on the Kamchatka peninsula they fester something similar, perhaps an unmentionable part of a brown bear buried in a peat bog for months. There are no available tyramine data on these. But if you have read this far without learning already that they are obviously to be avoided then

Fish sauces

See ‘Fermented Sauces’ above.

Malaysian budu and cincalok

Malaysian local appetisers ‘budu’ and ‘cincalok’ [102] tyramine up to 450 mg/kg.

Pizza

It depends what you put on it! It should be clear from the data in this monograph that almost all commercial pizzas are highly likely to be safe, as found by Shulman [162]. This is because they are most unlikely to use anything other than commercial processed cheese, or non-matured cheese types (e.g. mozzarella, which has no tyramine). Also, any salami type products on them are likely to be in small quantities, and also of the type that is low in tyramine. Pizza chains/franchises may change their cheese blend to stay abreast of cheese fashions, but for cost reasons are most unlikely to use large proportions of matured cheese that might have a higher tyramine content.

Gourmet pizzas may contain mature salami and cheese with higher tyramine concentrations, but the quantities are likely to be small, so the total tyramine load is unlikely to be problematic. The data herein should allow a reasonable estimation of the total amount of tyramine. Sensible caution is therefore appropriate with some “gourmet” pizzas and with large servings of some “commercial” pizzas.

Vegetables and fruits

Vegetables generally have total BA concentrations of only a few mg/kg and tyramine levels of about 0.2 mg/kg with a maximum of 1 mg/kg [163], but can these increase a little with spoilage ([see below re avocados](#)). As stated in ‘Key facts’ in normal sized portions all these things are safe.

Plants do produce an extra-ordinary range of amines and psycho-active alkaloids, many are part of the ancient battle whereby plants manipulate the behaviour of animals and enhance their own survival (e.g. think of opiates, cannabis, tannins, nicotine, atropine, hyoscyne, aperients & innumerable toxins). Many of these compounds are more common in a greater variety of plants than a casual reading of the literature would lead one to suppose. Their concentration varies greatly depending on many factors like plant variety, tissue type, stage of growth and attack by other organism etc.

Useful reviews are: [163-166], it has recently emerged that some of these compounds affect TAA receptors and TRP channels e.g. capsaicin, menthol [167, 168].

Avocado and banana and other oddities

There is one credible report of high BP after consumption of about six avocados, probably over-ripe [169]. There are no recent data on Tyr levels. The fact that there have been no further reports in more than thirty years indicates it is pretty difficult to ingest risky amounts via fruits.

A recent (2019) more extensive study of bananas, of various genotypes, including ‘plantain’, showed the amines tyramine, histamine, dopamine, serotonin, spermidine and spermine all **decreased** during the ripening in most genotypes [170]. Levels in the pulp were remarkably consistent across different genotypes (100 mg/kg) but varied considerably in the peel (max 300 mg/kg). This study must be considered as the most definitive data currently available concerning bananas, and the older data cited in previous versions of this monograph now takes a backseat.

Bananas can have significant dopamine, up to 400 mg/kg in the pulp, about 1,500 mg/kg in the skin [171], but probably little tyramine [166, 172], and seeing Borges for this and other amines updated.

The first report of dopamine was in 1958 [173]. Large amounts of banana (20 per day) may increase plasma dopamine concentrations [174]. This may be via release of endogenous DA, and or via L-DOPA or other precursors or releasers. So, although DA cannot cross the blood brain barrier brain (or only to a limited extent [175]), plasma DA may be elevated, and raised peripheral DA may raise BP by vasoconstriction. As with all plants, concentrations will vary greatly according to variety, part of plant, stage of growth, maturation, ripeness etc. and it is clear concentrations are much higher in the skin (1,000 mg/kg) than the pulp [171], at only 2 mg/kg [176] and see [177-179].

Banana might possibly inhibit the adsorption of medicinal L-DOPA [180, 181]. It is very unlikely that bananas in usual quantities would have any significant effect.

Paprika and green pepper appear to have higher tyramine contents 286 and 141.5 mg/kg of dry weight, respectively [164].

There is always something new. Ever heard of ‘fermented minced pepper’. Well evidently it is enjoying rising popularity in China! It may well have slightly elevation tyramine levels [182], so do not eat it by the jar-full.

In summary, it would seem normal servings of fresh vegetables, fruits etc. are unlikely to have any serious adverse effects via histamine, tyramine, or L-dopa (that includes broad-beans, aka fava beans, and related species).

Nevertheless, interactions are sometimes noticeable and there is much yet to learn about the psycho-active contents of plant derived foods. One interesting recent reaction [personal communication] involved a reliably documented alteration of BP associated with consumption of quince paste, for which there would appear to be a possible explanation, since it has been claimed to contain a constituent that acts as a dopamine re-uptake inhibitor [183]. This finding needs to be replicated, especially because the probity of much Chinese research is doubtful [184].

Spinach

Tyramine in spinach [185] was < 5 mg/kg, but histamine can be higher ~ 50 – 100 mg/kg.

Fava/faba beans

Fava beans (*Vicia Faba*, aka broad beans) have tyramine at about 10 mg/kg [163], & L-DOPA, but at low concentrations, which is probably not sufficient to have any effect in normal portions. See ‘L-DOPA’.

L-DOPA

Dopamine (DA) is present in many plants and plays a role in repelling pathogens. It is the precursor of the quinones that cause browning when they polymerise into melanin (e.g. bananas, avocados). Some legumes contain significant amounts of L-DOPA in some tissues, at some stages of growth, including *Vicia faba* L. varieties (aka fava beans, broad beans) and *Mucuna pruriens* (Cowhage, itching powder) [186-191]. Varieties of these plants are being genetically engineered to try to find a suitable

dietary source for L-DOPA because it may be better than pharmaceutical L-DOPA (better absorption, more stable plasma concentrations). Various preparations are being sold on the internet. A search for ‘mucuna aphrodisiac’ or ‘mucuna parkinson’ returns many thousands of hits.

Maximum concentrations of 10–20 mg/g (dry weight) have been found in *Vicia faba* [186], equivalent to a wet weight concentration of approximately 100 mg/kg. However, the edible beans are lower.

Since L-DOPA is a dopamine precursor, not a releaser, i.e. not an indirectly acting sympathomimetic like amphetamine is, it is likely to have an effect more analogous to L-tryptophan with MAOIs (i.e. moderate potentiation only). L-tryptophan does not cause serious problems with serotonin toxicity, and nor would one expect L-DOPA to do so with BP.

Despite the warnings on interactions with medicinal L-DOPA, early papers were, [192-194]. The evidence for serious hypertension (see below for discussion) with L-DOPA and MAOIs seems inconclusive.

Such amounts of L-DOPA may potentiate or precipitate moderate BP increases, but, in my opinion, it is unlikely that a seriously risky BP elevation would result.

Pickling and preserving

Preservation, mostly of vegetables, using the acidic properties of natural acids, mostly acetic and lactic acid, is widespread and usually involves no fermentation, just the addition of vinegar (acetic acid), as in typical pickled onions. However, other pickled preparations involve a bacterial lactic acid fermentation process, such as sauerkraut and kimchi, see below. It is these fermentation processes which can give rise to small amounts of tyramine. Naturally occurring fermentation, without the use of starter cultures (see above) tends to produce more contaminant biogenic amines, including tyramine.

Olives, capers, caper berries

Preparation of olives may involve bacterial lactic acid fermentation, tyramine levels in olives, and capers are very low [195, 196].

Sauerkraut

Sauerkraut is made by lacto-fermentation, as are kimchi & traditional pickled cucumbers. These keep for several months, unrefrigerated.

Sauerkraut: review [108], more than 100 samples from 7 countries, almost all tyramine < 200 mg/kg, but a couple from Czech Rep. were 400 – 900 mg/kg.

Tyramine concentration was 50 mg/kg in one canned sauerkraut, other samples < 12 mg/kg.

Korean ‘kimchi’ cabbage average tyramine 50 mg/kg, max 120 mg/kg [82].

Lavizzari [165, 166]: Spinach tyramine 2 mg/kg. Histamine concentrations were 100 mg/kg.

Kosson [197] found insignificant levels of tyramine.

Chocolate

Chocolate sometimes does involve a short fermentation stage. Somewhat variable concentrations of amines have been reported, mostly very low, and inconsequential — unless *large* quantities are consumed (i.e. more than 100 grams).

A few recent papers have added data on dozens of samples of cacao powder, chocolate (white, milk, dark) and syrup, see especially [198, 199], none of which exceeded 35 mg/kg, most being < 10 mg/kg.

These other results are in the same range: Pastore found 2 mg/kg for tyramine [200]; Lavizzari [166] found concentrations of tyramine of 0.3 mg/kg; Baker [201], powdered cacao: tyramine 3 mg/kg, chocolate < 1 mg/kg & Granvogl [202-205].

Health supplements

Such substances can contain all sorts of ‘illegal’ additives, some potentially injurious, and most of them useless. The commonest adulterants are SRIs (like sibutramine — danger of ST), steroids, stimulants, and sildenafil (Viagra). They should list the main ingredients, if they do not, then they should not be used. If they do, then do not use them if they contain tyramine at levels that would be injurious, i.e. more than 5–10 mg per portion or ‘dose’ [206].

Other non-serious interactions

Many plant-derived substances (alkaloids), e.g. ‘herbs’ and ‘foods’ like coffee, and tea contain various compounds that act as ‘drugs’, stimulants like caffeine, 2-phenylethylamine, methylamine, trimethylamine (see Strolin Benedetti & Tipton [207]). These affect everyone but may have an exaggerated effect in those taking various sorts of antidepressant drugs, including MAOIs; they should be taken in moderation and avoided if they precipitate symptoms such as tremor, anxiety, jitteriness, palpitations, tachycardia, agitation, or poor sleep.

Some tyramine champions

One soy sauce clocked in at 6,000 mg/kg [95], does one smell a rat there?

An old cheddar cheese measurement from the 1950s 3,700 mg/kg [53].

An Italian goat cheese at ~ 2,000 mg/kg [208]

And, there is a French cheese called ‘crotte du diable’ (translates as ‘Devil’s turds’), and various rotten-fish brews (best consumed on isolated Scandinavian mountain tops), that one presumes would be contestants, but I was unable to find any data. Would any lab technician be brave enough to endure them?

For an introduction to some other strong-smelling foods see Andrew Zimmern:

<http://www.openjourney.com/article/18-stinky-foods-around-the-world-41.html>

Holidays

Some holiday destinations will require heightened awareness of food hygiene issues, in “Biogenic amine contents in selected Egyptian fermented foods as determined by ion-exchange chromatography” Rabie found levels of 2,000 mg/kg in cheese and

fermented sausage [209], then there is fermented Yak milk [83], and Icelandic fish-dish called Hákarl (fermented shark meat).

Wine, spirits and beer

A meal without wine is like a day without sunshine.
Anthelme Brillat-Savarin

Wine and beer in moderation (two drinks in 2 hours) are definitely safe (as far as tyramine is concerned). Modern hygienic production methods for beer have made tyramine concentrations > 10 mg/L rare (there is now extensive regulation and documentation of this, see below for details). Home-made wines or beers may be risky. Bottled beer is safe if pasteurised; a little caution is warranted with 'live' beers which may be available from 'boutique' producers. They can be distinguished by the sediment (of dead yeast) in the bottom and they are cloudy if shaken.

Modern commercial wines do not contain significant tyramine.

Tyramine in liquids taken on an empty stomach should be regarded as a special case, because tyramine will be absorbed much more rapidly [210, 211], so amounts of tyramine of one third of the figures given above may evoke a reaction. One small (330 ml) glass of some 'live' beers could, in *rare* instances, have about 10 mg of tyramine; this is sufficient to cause a reaction in a minority of people, when taken on an empty stomach, e.g. see [36, 212].

Wines

Here with a loaf of bread beneath the bough,
A flask of wine, a book of verse – and thou
Omar Khayyam

Wine can *very rarely* contain significant concentrations of tyramine (> 10 mg/L).

Recent major reviews have covered many hundreds of different wines of all types: almost all have had tyramine levels of less than < 5 mg/L [213-222].

Aged wines, all tyramine < 5 mg/L [223].

Thirty different wines, including aged fortified wines (Port and Madeira), max tyramine 5 mg/L [224]. Wines 200 samples, histamine average 1.2 mg/L [225] and 300 samples max tyramine < 5 mg/L [226, 227].

USA wines, max tyramine 3 mg/L [217].

Marcobal, 61 different Spanish wines including aged Rioja Gran Reserva wines [228]: Tyramine range 0–11.32 mg/L, Average 1.40 ± 2.35 mg/L. Only 34 of 61 wines had *detectable* tyramine.

However, Preti et al. found 8 of 60 (personnel communication) Italian wines tested recently had tyramine levels > 10 mg/L [229], none of these were chiantis!

The repetition of the notion that Chianti, uniquely amongst wines, contains significant concentrations of tyramine [70], illustrates, so it seems, how easy it is to be careless about the relevance and reliability of sources of information. The chianti error was countered long ago [230]. The most likely explanation for these anomalies is that in the past many of these wines were made by farmers with little knowledge of wine or

fermentation techniques. Hygiene practices were poor; it is only in the last 20 years or so that Italian winemaking has reached a modern standard, in most places.

Vinegars

Ordinary vinegars low tyramine, but: Chinese rice wine vinegar (old) 400 mg/L, Sherry vinegar 15 mg/L, Italian Balsamic ~ 15 mg/L [231].

Beers

Standards, and awareness of brewing hygiene issues, have increased since some of the older results, but caution is still warranted: it would seem likely that most, but not all, standard commercial and modern beers all over the world will be safe (< 10 mg/L) in moderation; some low volume ‘artisan’ and ‘boutique’ ones are a little more likely to be risky. Beers made using natural yeasts (spontaneous fermentation) rather than starter cultures, are more likely to have contaminants and therefore high tyramine. This is an observation echoed throughout this monograph with all types of ‘fermentation’, whether with cereals or sausages. Some examples are high enough to be risky, especially if beer is drunk on a more-or-less empty stomach, when it will be absorbed more rapidly.

It is established that the presence of tyramine is indicative of bacterial contamination and less than ideal hygiene practice.

A review by Kalac [232-234], “195 samples of bottled or canned beers were purchased from commercial outlets in Germany, Austria, Belgium, Bulgaria, Czech Republic, Denmark, Spain, France, Great Britain, Greece, The Netherlands, Ireland, Italy, Portugal, Switzerland, and the former Yugoslavia”. They found a great majority were low (2 – 8 mg/L, mean 7), but a few are up to 30 – 50 mg/L, with a maximum of 70 mg/L.

Bunka more recently reviewed 114 samples of beer from 28 breweries in the Czech Republic which were “monitored at their purchase and at the end of their best-before period” [235]. Tyramine was < 10mg/L in 51 samples, between 10 and 50mg/L in 21 samples and 100 mg/L in 5 samples.

Pradenas et al. in Chile assayed over 100 samples and found 99% of 316 beer samples were no more than 2 mg/L, one was 6 mg/L [236].

Tang [237]:18 beers all brewed in China, some European under licence, values mostly tyramine 3 – 5 (max 7) mg/L.

Spanish beer: tyramine < 2 mg/L [238]; 17 samples mean tyramine 5 mg/kg; 55 samples mean 7 mg/L, max 47 mg/L. Europe 48 samples max 6 mg/L [239, 240].

16 European countries, 195 samples, mean tyramine 6.5 mg/L max 67.5 mg/L [241].

17 domestic Turkish and 13 imported beers [242], all were tyramine < 2 mg/L.

So, a great majority are low and safe, but if your favourite tippie is a ‘micro-brewery’, open fermented or ‘live’ or something exotic like Belgian lambic, watch it! Test it out very carefully before swigging too much!

Ken Shulman’s group [243] looked at a total of **98 beer samples** (79 different brands of beer) in 1994:

'All of the bottled beers analysed had safe tyramine concentrations (< or = 10 mg/liter; range, 0 to 3.16 mg/liter) and, thus, do not require restriction in patients receiving MAOIs. Therefore, the consumption of canned or bottled beer, including dealcoholized beer, in moderation (fewer than four bottles or cans; 1.5 litres within a 4-hour period) appears to be safe and does not require restriction in patients receiving MAOIs. Only 4 of 98 beer samples studied were found to have a dangerous (> 10 mg/liter) tyramine concentration, one of which was the index beer. The tyramine concentration in these four beers ranged from 26.34 to 112.91 mg/liter. All four of these beers were tap beers produced by bottom fermentation (lagers) and brewed by a secondary fermentation process. ... Therefore, to err on the side of caution, it is recommended that patients on irreversible MAOIs avoid beers on tap'.

This was an influential paper; subsequent knowledge suggests a slight modification of their conclusions.

Belgian beers especially can have high tyramine. Loret et al. [244], considered a large number of these Belgian beers: the types covered four different brewing processes; low or bottom fermentation (LF, 18 samples), top fermentation (TF, 36 samples), top fermentation followed by a secondary fermentation in bottle (TF+ BSF, 184 samples), and spontaneous fermentation (SF, 42 samples).

They found 21 samples out of 220 that exceeded 10 mg/L of either histamine or tyramine, these 21 had a mean tyramine of 28 mg/L, and the maximum was nearly 70 mg/L. They developed a "Beer biogenic amine index" (BAI) that would allow assessment of the quality of the production process. Since the work was financed in part by the Belgian Brewer Confederation, we may assume they are trying to improve things because of EC regulations and a recommended limit of tyramine 10 mg/L.

Belgian Lambic beer is an old style (see Wikipedia for information) allowed to spontaneously ferment with wild airborne yeasts and then aged for 1 – 3 years, breweries locate their open fermenters in well-ventilated attic roofs. The general category is spontaneously fermented beers (SF beers) which are obviously likely to have more tyramine (because they have more 'contaminant' organisms).

One more recent assay of SF Belgian beer found only 20 mg/L of tyramine, which may well reflect improved standards [244]. Gueuze is an aged unflavoured Lambic style. This is a good illustration of why dirty farmhouse styles of anything are more likely to have contaminant strains that have decarboxylase activity, and thus potential for tyramine production, especially if a rat/sparrow/cockroach falls into the open fermenter.

MAOIs and scombroidosis (histamine fish poisoning)

The anti-tuberculosis drug isoniazid (INH) is closely related structurally and pharmacologically to phenelzine, but not related to tranlycypromine. INH is capable of inhibiting one of the other amine oxidase enzymes, the one which is largely responsible for breaking down histamine. The result of this is increased sensitivity to any histamine ingested in food by patients on INH and many histamine reactions have been described [245-252]. The potency of phenelzine (but not TCP) for these effects is probably similar to isoniazid, and the blood and tissue concentrations reached in the system are also probably similar. However, there have been no reports of definite histamine reactions involving phenelzine: nevertheless, it is probable that phenelzine does increase people's sensitivity to histamine. The seminal early work by Blackwell and Marley is still well worth reading, and indeed, they presciently predicted

sensitivity to histamine as discussed in this MS — they were indeed way ahead of their time [48].

Bearing in mind that foods that accumulate tyramine, like cheeses, may also have elevated histamine concentrations, this may be of relevance to patients taking phenelzine.

Symptoms of histamine poisoning are lowered BP, headache, palpitations, skin flushing, nausea, vomiting, and pruritus (itching).

The symptoms of histamine poisoning relate especially to effects on blood vessels, cell permeability and smooth muscles, and include headache, nasal secretion, bronchospasm, tachycardia, extra-systoles, hypotension, edema (eyelids), urticaria, pruritus, flushing and asthma [253, 254]. Serum tryptase concentrations may help to distinguish allergic symptoms from scombroidosis [255].

It is inevitable that some instances of BA poisoning will exhibit mixed symptoms of both histamine and tyramine effects, especially in people taking hydrazine drugs like carbidopa, isoniazid and phenelzine, and as above, Blackwell and Marley discussed this, but it does not seem that anyone has read, or at least remembered, what they wrote — maybe we might stop a moment and raise our glasses for a toast in their memory.

Marley went on to write more seminal papers on MAOIs that are important vis-a-vis ST [256-261], those were also ‘over-looked’ by almost all writers: so raise your glass again.

Part 2 Drug interactions

The place of MAOIs in treatment

A brief survey about the place of MAOIs in modern practice provides perspective and reveals the disproportionate influence on doctors' prescribing practices of 'promotion' in its various and often deceitful guises. The pressure to influence doctors to prescribe new drugs has been driven by pharmaceutical companies and has been hugely successful. It is a triumph of greed and profit, barely restrained by ethics. The commercialism and advertising that now dominate science, to such a disproportionate extent, are regarded by many as a great problem. This has been dubbed 'McScience' and it is the new reality, as the erstwhile Lancet editor warned us a decade ago [262, 263]:

'Journals have devolved into information laundering operations for the pharmaceutical industry'.

The incredibly low rate of prescription of MAOIs is starkly incongruent with the fact that they are recommended and endorsed by many reviewers and in all recent guidelines about the treatment of depression [2, 57, 264-277].

Yet only a tiny fraction of specialists ever use MAOIs [57, 274, 278, 279], despite opinion and evidence of their superior effectiveness for various groups of patients [13, 272, 276, 280-283].

My international expert group on MAOIs has just published an editorial statement on this subject which will hopefully be of some influence [10].

MAOIs: Interactions with other drugs

Science must begin with myths, then progress to the criticism of myths.
Karl Popper

Myth: MAOIs have many problematic interactions with other drugs.

Yet there are no pharmaco-kinetic interactions and just two pharmaco-dynamic interactions: SRIs and releasers.

SRIs are easy to identify and avoid. Releasers rarely used therapeutically and rarely cause reactions sufficiently severe to be high risk.

And doctors have been convinced that is difficult to cope with? I hardly think so.

The requirement, a simple requirement, is to learn which drugs are SRIs and which are releasers. Then it is all plain sailing.

It is helpful to understand why this text, and my review papers, contradict what is said in standard textbooks and other similar sources (e.g. Physicians' Desk Reference, British National Formulary, Australian Medicines Handbook etc.). First, many of them are just plain wrong. Second, such publications cover a wide field as concisely as possible and therefore abbreviate and generalise to an extent that does not allow detailed evaluations. For example, such sources lump all tricyclic antidepressants together as being contraindicated with MAOIs. Such texts have insufficient space to discuss more precise considerations detailed in review papers and in this monograph. The view that the average doctor cannot understand such subtleties may also be a factor.

MAOI interactions are now clearly understood***, they are reliably predictable, and they are straightforward to avoid. There is no room for a lengthy discussion here, but readers may note that I have published widely concerning both pharmacokinetic and pharmacodynamic interactions, and the cytochrome P-450 characteristics, of most psychotropic drugs. Consult those papers if you wish to have more understanding of this subject. They provide the background knowledge for understanding these interactions. See particularly my review, 'CNS toxicity involving methylene blue: the exemplar for understanding and predicting drug interactions that precipitate serotonin toxicity' [4], which is my most recent summary of what needs to be understood in order to be confident about avoiding ST.

*** As a matter of historical record and to recall the admonitions about learning from history — both the 1964 JAMA editorial [284], which used the word 'hysterical' (with which I agree) and Blackwell [50] opined that the American reaction of taking Parnate off the market (for six months in 1964 — the British did not follow suite) was way 'over the top'. Blackwell correctly pointed out that the 'cheese' reaction should have been anticipated from pre-existing research (cf. Marley); and I have said the same about the imipramine/MAOI interaction (i.e. ST), it took about 30 years before that was properly understood. Psychiatrists have a poor record as students of pharmacology. It was ever thus. It is something about which psychiatrists should be ashamed. It is poor knowledge, combined with a therapeutically pusillanimous attitude, that is largely responsible for the enduring negative/nervous attitude to MAOI use (it is just 'too difficult' for most of them).

For other aspects of interactions, or rather, lack of them, see also: [2, 5-7, 19, 20, 285, 286]. There is an updated summary about the lack of interactions between MAOIs and TCAs [here](#).

Non-therapeutic/illicit drugs

This commentary deals with licit therapeutic drugs.

MAOIs (including moclobemide) combined with ecstasy (MDMA) carries a high risk of fatal ST.

Those requiring information about non-therapeutic/illicit drugs are advised to be wary 1) there is a lot of mis-information in medical texts and on the net. Some interact potently with MAOIs, because many of them are potent releasers of serotonin, dopamine and or noradrenaline [287]: e.g. the interaction of moclobemide and MDMA is predictably toxic (causing fatal ST) and has caused a number of tragedies [288, 289]. Note that combinations of releasers with re-uptake inhibitors will result in diminished effects/efficacy: so, for example, SSRIs will diminish the effects of 3,4-methylenedioxy-methamphetamine (MDMA, ecstasy). For further explanation about this see my review [4].

The following papers contain information and further references about 'designer' and novel psychoactive substances [287, 290-293].

Medically, such possible interactions are only likely to be seen as presentations to emergency departments and are unlikely to be relevant to usual therapeutic practice.

MAOIs are not 'dangerous'

It is common to hear and read of MAOIs being described as 'dangerous'. That is neither logical nor reasonable.

Being ill with unresolved or poorly treated depression is much more risky than taking MAOIs because not only is the life-time risk of death by suicide around 10%, but also the Standardised Mortality Ratio SMR (which includes death from other causes than

just suicide, such as heart disease which is increased in those suffering depression) is as much as 10 – 30 times elevated [294-305].

The view has been well argued by the eminent medical historian, Edward Shorter, that the dangerousness idea was encouraged and spread by pharmaceutical companies extolling the virtues of newer drugs [306], and that necessarily involves exaggerating the disadvantages of previously existing drugs. A [white paper by White and Simpson](#) for the ACNP (adopted as an official ‘position statement’ in 1981) also used the word ‘superstition’ to describe the concern about the risks of MAOIs.

Tranlycypromine has no clinically relevant pharmaco-kinetic interactions and phenelzine has almost none, certainly none that are clinically significant [2], which makes them much better, certainly in this respect, than most of the SSRIs and some other ‘new’ drugs.

The potentially risky interactions with MAOIs are the pharmaco-dynamic ones, the more likely and serious being:

1. Serotonin syndrome, caused by SRIs + MAOIs

And the other, less-frequently

2. Blood pressure elevation, [much less risky](#), caused by tyramine in food, or by the releasers like ephedrine.

Interactions with SRI anti-depressant drugs and other SRIs

Any drug that works as a serotonin reuptake inhibitor (SRI), not just the SRI anti-depressant drugs, but including the SNRIs and a few other drugs that act as SRIs (even if not marketed as such) produces a high and predictable risk (possibly fatal) of ST, if combined with a usual *therapeutic* dose of an MAOI (which includes **reversible inhibitors of monoamine oxidase A (RIMAs)** like moclobemide [3, 285, 307, 308]. ST is not an idiosyncratic interaction; it is a predictable dose-related toxic interaction.

In the last 20 years or so ST has led to a small number of deaths, entirely avoidable and many due to the ignorance of doctors about this interaction. A pressor response to excess tyramine has led to few, if any, deaths over this same period.

The category of serotonin reuptake inhibitor drugs includes:

- 1) All SRI and SNRI anti-depressant drugs: sertraline, fluoxetine, paroxetine, fluvoxamine, citalopram, escitalopram, vortioxetine, clomipramine and imipramine* and chlorpheniramine (aka chlorphenamine) and brompheniramine, or SNRIs: milnacipran, levomilnacipran, venlafaxine, desvenlafaxine, duloxetine or sibutramine.

* It is usually stated that all TCAs pose a risk, but that is definitely not correct, it is only clomipramine and imipramine that are sufficiently potent as serotonin reuptake inhibitors to precipitate ST; all other TCAs like nortriptyline, amitriptyline, trimipramine, dothiepin, doxepin, desipramine, protriptyline, are quite safe (as are selective NRIs like reboxetine and atomoxetine).

NB There have been one or two other ‘odd’ ‘SRI’ drugs like SNRI anti-depressant sibutramine [309, 310] that were (eventually) marketed for other indications (appetite suppressant), but sibutramine has been withdrawn world-wide [311, 312].

For a [database of withdrawn drugs](#), see [313]

The anti-histamines brompheniramine and chlorpheniramine (aka chlorphenamine), should be avoided because they have significant SRI potency [314-324]. Other anti-

histamines are safe, but better SERT affinity data would be reassuring. NB the current generation anti-histamines do not cross the BBB, so cannot contribute to ST.

All of these old tricyclics came from the same pool and some were inappropriately classified as antidepressants, such as doxepin, and some inappropriately as antihistamines, e.g. chlorphenamine. Indeed, it would be logical, once more accurate data is available, to reassign them to different pharmacological groups based on their actual properties (cf. discussions elsewhere about ‘neuroscience-based nomenclature’ of drugs). Meanwhile, it is useful to remember that doxepin (classed as a TCA) is the most potent antihistamine currently on the world market, and it definitely does not have any significant SERT affinity, and definitely does not cause significant ST.

2) Some narcotic analgesics, because some of them also act as SRIs: meperidine (aka pethidine) and tramadol [20] and less often used dextro-propoxyphene and dextro-methorphan. They are only weak SRIs, so therapeutic doses will rarely cause ST: it is usually only after larger and repeated doses that serious ST might occur.

All non-narcotic analgesics are safe to take with MAOIs: aspirin and paracetamol and all the NSAIDs etc.

Releasers (indirectly acting sympatho-mimetics ISAs)

One or two recent papers about the mechanisms of action of MAOIs and amphetamine at the molecular level suggest why the combination of amphetamine (with MAOIs) is not unduly risky as has been (mis)stated for so long. **Care and experience are required** but it can be done safely, although small increases in dose do sometimes seem to have disproportionate effects. The advice of ‘start low go slow’ is especially applicable and it should be appreciated that it is likely that BP will be increased to a modest but significant degree. This may be relevant and problematic in some patients but may be a distinct advantage in patients who suffer disabling postural hypotension. If hypotension is not a problem, then modafinil may be useful alternative.

Amphetamine is a potent DA and NA releaser (in former terminology, an ISA) at low nano-molar (10^{-9}) concentrations. There is still uncertainty about its exact mechanisms of action and just how it interacts with the monoamine transporters etc. It acts as a competitive inhibitor (for NAT & DAT) and has actions in the pre-synaptic cytoplasm, at the vesicular monoamine transporter (VMAT) and TAR1 receptors. The latest understanding of this is complex and beyond the scope of this review. Further details are in: [325, 326]. Those using these combinations may wish to study these references to understand more about ‘non-exocytotic release’ that does not require any neuronal activity (*viz.* nerve impulse) to trigger it, and how they inhibit reuptake in a competitive manner and more.

Unsurprisingly, considering the multiple sites at which drugs classed as amphetamines affect the neurotransmitter systems, there are marked differences between closely related drugs: for instance, methylphenidate (which is classified as an amphetamine) is a DA re-uptake inhibitor, and not a releaser. It produces no risk of ST, nor a pressor response.

As Paracelsus stated, ‘*the dose makes the poison*’ and that may be particularly applicable to amphetamine. Releasers are capable of increasing intra-synaptic transmitter concentrations by more than 1,000-fold, compared to a maximum closer to 10-fold

with reuptake inhibitors [326] — cf. [7] re such mechanisms of interactions involving RIs, releasers and MAOIs.

Amphetamine causes NA increases of a lesser magnitude (400–450% of baseline) compared to dopamine (700–1500% of baseline). This suggests that used carefully the risk of precipitating hypertension is low (as practical experience indicates, see Israel for a recent report and review [327]). The advent of lisdexamfetamine may now add another layer of safety because its slow conversion to the active form (d-amphetamine) occurs in red blood cells by rate-limited enzymatic hydrolysis. This means the time to T_{max} is rather longer and peak levels are lower, about half [328]. It also has a low potential for cytochrome P450 interactions [329, 330]. Not only that, but also the inter- and intra-subject plasma levels are much less variable which produces a ‘smoother’ and more predictable response [331]. An interesting example of the usefulness of a pro-drug. It is to be expected that this combination (with MAOIs) will be even safer than previous preparations [327, 329, 332-336].

Ephedrine is rather less potent than amphetamine [337-339]. Pseudoephedrine is much less potent than ephedrine.

Ephedrine, and to a lesser degree, pseudoephedrine, are the archetypal drugs of concern, and are still available for use in some countries, whereas in most they have been replaced by oxymetazoline (which does not interact with MAOIs). Previously they were components of cough and cold remedies. Reactions with ephedrine may be severe but are less severe with pseudo-ephedrine.

Adrenaline (epinephrine) and noradrenaline (norepinephrine) are, obviously, (because they are the body’s neurotransmitters that act at these receptors) direct post-synaptic agonists and therefore do not cause any problematic interaction with MAOIs. Equivocation about that has been evinced repeatedly over the years in most standard texts and has caused mistreatment of patients e.g. [340], yet the lack of an interaction was established at the dawn of modern pharmacology by researchers whose names are prominent in history (Gaddum and Brodie, among others), early papers being [341-343]. That work has been forgotten. It is TCAs that have a more pronounced interaction with adrenaline, ironically, I cannot recall anyone getting too worried about that.

There is now quite a lot of accumulated experience of the concurrent administration of MAOIs and amphetamine for therapeutic purposes in depression. It is safe when done carefully. Early concerns about frequent hypertension have not materialized and recent clinical reviews indicate judicious use is safe [344, 345]. Since amphetamine is substantially more potent than ephedrine it would seem, by extension, that concerns over this drug may also have been over-rated. If taken in supra-therapeutic doses or overdose the situation may be different.

Traditionally concern about interactions has centered around cough and cold remedies and nasal decongestants because of early confused reports in the 1960s, e.g. [346, 347] and because they may contain both SRIs (e.g. chlorpheniramine (aka chlorphenamine), dextromethorphan and releasers like ephedrine). Note that until the 1990s, and in some reports beyond, there was a failure to understand the toxidromic distinction between a risky pressor response and ST. That failure has caused much confusion. The unrecognised irony was, until my 1998 review, it was not recognised that the chlorphenamine component of such over-the-counter (OTC) remedies is an SRI, and therefore a potential problem for precipitating ST. Indeed, as

I noted, chlorphenamine was a possible, but unrecognized, contributor to the death of poor Libby Zion in a much, but inaccurately, commented on case [348-350].

One may also note here another classic case of pharmacological misunderstanding. It did not attain notoriety, the doctors were lucky because the patient was on a sub-therapeutic dose of phenelzine (only 15 mg daily) and therefore did not get ST [340]. These doctors misguidedly eschewed adrenaline to treat an anaphylactic reaction because their patient was on phenelzine, so instead gave intravenous chlorphenamine. In view of the text above no further explanation of this, fortunately comical, error should be required.

Chlorphenamine, in the usually recommended doses of 10 – 20 mg IV, and up to 40 mg in 24 hrs., is almost certain to attain sufficient blockade of SERT to precipitate ST in combination with MAOIs. The warning with it, below, is a good example of the kind of unhelpful mis-information that is still in ‘official’ texts:

‘The anticholinergic properties of chlorphenamine are intensified by monoamine oxidase inhibitors (MAOIs). Chlorphenamine injection is therefore contraindicated in patients who have been treated with MAOIs within the last fourteen days.’

Incredibly, not a word about SERT inhibition and the probability of ST. Incidentally, I wrote to the regulatory authorities about that years ago. My communication was ignored. Some culpability there one might think.

Typical constituents of available cough and cold remedies and nasal decongestants: phenylephrine, pseudoephedrine, both weak releasers and now mostly replaced, or phenylpropranolamine now completely withdrawn. The usual constituents of cough and cold remedies now are the MAOI-safe alternatives: oxymetazoline, xylometazoline and ipratropium bromide.

Herbal preparations: ephedrine is in various plant species, e.g. *Ephedra sinica* (Ma Huang).

As Rothman states, ‘Historically, it has been difficult to distinguish whether drugs act as reuptake inhibitors or substrate-type releasers using simple test-tube assays.’ But it seems now established that amphetamine is a moderately potent NE and DA releaser, but a weak 5-HT releaser [337-339].

Therefore, over-the-counter drugs are hardly a problem now, because even pseudoephedrine has been taken off the market (at least, in many western countries).

The commonest ‘non-releaser’ nasal decongestant is oxymetazoline, which is an adrenergic alpha 2 agonist: it has no interaction with MAOIs and is not a problem.

Directly acting agonists, such as midodrine and adrenaline itself, are not a problem with MAOIs, because there is no potentiation, something that was established over half a century ago. There is an additive effect so dose adjustment will be required when they are used together.

In summary: releasers are almost a problem of the past, and, except for ephedrine itself, are unlikely to cause severe reactions in normal moderate therapeutic use.

Anti-psychotic drugs

All available anti-psychotic drugs have, until recently, been safe with MAOIs. However, one newer so-called atypical, ziprasidone (Zeldox), possess SRI potency

and has been implicated in a case of definite ST in combination with an MAOI [351], it is therefore contra-indicated with MAOIs.

Triptans

There is no risk of ST with triptans. The FDA have issued various misconceived warnings, particularly about triptans and serotonin toxicity: see Gillman [5]. There has been no subsequent rebuttal of the conclusions in that review, that there is no risk of ST with triptans, either by the FDA or by anyone else. Other reviews and comments concur with my conclusion [352-355].

Anaesthesia

Myth: One cannot give an anaesthetic without ceasing MAOIs first.

The main issue in most operations, and post-operative periods, is the avoidance of analgesics that act as SRIs, viz. meperidine (pethidine), tramadol, tapentadol, dextromeporphan and pentazocine. Other opioids like codeine, oxycodone, morphine, buprenorphine, fentanyl, are safe (see [20]).

The idea that an anaesthetic cannot be given without first ceasing MAOIs is yet another of the deeply embedded and ill-founded concerns that one encounters. Sadly, it is not inconsequential, because poorly informed careless surgeons (some of whom might struggle to spell ‘pharmacology’) may tell patients due for elective surgery to cease treatment, sometimes without being aware of their history. I have had experience of suicides from relapse of depression as a direct result of such ill-advised cessation of treatment. Therefore, my [disparaging view of those surgeons](#) who are too careless, ignorant, and arrogant to ask advice, or even inform the prescribing specialist that they have ceased the treatment, will be understandable to some.

In ‘uncomplicated’ anaesthesia, apart from avoiding any use of narcotic analgesics with SRI potency, there are no major problems or interactions. The preponderance of published opinion has supported that view for some time [356-364]. There is further irony in the tendency for people to overemphasise the risks of MAOIs, but forget the risks associated with SSRIs (bleeding) and TCAs.

For ‘major’ operations that might require treatment to raise or lower blood pressure there are some adjustments of dosage and agents that may be required, but there are no major obstacles. For instance, the hypotensive effect of MAOIs may mean that intra-operative hypotensive measures may be potentiated, and accordingly doses of such drugs may need to be lower. On the other hand, if vasopressor agents are required then directly acting alpha agonists may have their effects potentiated, that means norepinephrine, epinephrine and phenylephrine doses may need to be slightly lower when used in patients on MAOIs. Since ephedrine has releaser potency it is best avoided.

Stopping and swapping, easier than supposed

On ceasing SRI-type antidepressants to start MAOIs, washout intervals varying between one and five weeks may be required. No washout is needed for non-SRI-type drugs. The rule of thumb is to allow 5 half-lives to elapse*, which is about one week for many of these drugs).

See, for a table of half-lives: <http://www.health.harvard.edu/diseases-and-conditions/going-off-antidepressants>

No washout is required for TCAs (other than clomipramine and imipramine), or mirtazapine, mianserin, bupropion, trazodone, reboxetine or atomoxetine, because they are safe taken together with MAOIs (i.e. anything at all is safe, except an SRI).

*Fluoxetine (via its metabolite norfluoxetine) has an elimination half-life in some people of up to two weeks (so it can take up to 10 weeks to get out of the system).

In practice 5 half-lives is a conservative approach. Most drugs will have lost sufficient SRI activity after two half-lives to allow cautious introduction of an MAOI- providing the patient can be observed for early signs of serotonin-related signs. Such signs are: 1) specific; tremor, hyperreflexia and clonus; less specific; GI overactivity, mydriasis, sweating, anxiety, restlessness, overactivity.

Should such symptoms be apparent it is simple enough to withhold the MAOI for another day or two and then try again. ST is a dose-related phenomenon and the emergence of more pronounced serotonin-related side effects is not a cause for immediate panic.

It is commonly thought and stated that it is problematic to change from one drug to another when one of them is an MAOI. In practice this is not a difficulty, once it is appreciated that almost anything, except an SRI, can be safely co-administered. The TCA nortriptyline is probably the best and most flexible candidate to fulfill a 'bridging' role because of its favourable pharmacological characteristics (no active metabolites, minimal pharmaco-kinetic interaction potential, medium length of half-life, mild sedative, and low anti-muscarinic potency).

For instance, if one is changing between an MAOI and venlafaxine, which can have quite marked withdrawal symptoms, co-administering nortriptyline prior to reducing or ceasing the venlafaxine can both reduce withdrawal symptoms and act as a 'bridge' prior to the initiation of the MAOI. Exactly the same process works in reverse where nortriptyline can be added to a pre-existing MAOI, the MAOI is then stopped and most subsequent treatments can then be initiated with ease.

*One could use mirtazapine, doxepin, amitriptyline, quetiapine, (but not ziprasidone), mood stabilizers: in short, anything that is not an SRI. How difficult is that!

Swapping from one MAOI to another MAOI

The requirement or desire to swap from one MAOI to another MAOI is something that will be a uncommon occurrence. Furthermore, it will be an urgent need even more rarely. It may be indicated, for instance because of the excessive weight gain, sexual dysfunction, or oedema, that occur with phenelzine. Because opinion exists in the literature suggesting dove-tailing, or a direct swap, is a potentially risky thing to do, some discussion about this is educative.

Insofar as I have been able to trace the original texts which postulate these sorts of dangers, I have one comment to make. Some are written by clinicians who have an incomplete understanding of pharmacology, or toxicology, and they contain errors of fact. Such texts are often books, most of which are not peer reviewed.

First, there is no known mechanism of interaction that would cause a problem. Neither is there a basis for postulating a pharmaco-kinetic interaction. Nor is there a basis for a major pharmaco-dynamic interaction. Which leaves a mystery, or possibly a phantom?

Second, my extensive experience analysing hundreds of case reports of ST illustrates clearly that the overwhelming majority of case reports are misleading*, and they often involve supposed interactions that have no known basis, in fact or theory. To make

decisions based on such reports has repeatedly proved to be inappropriate and the resultant actions have in some cases had serious negative consequences.

*This is why many reputable journals decline to publish case reports [286].

The literature is replete with inappropriate and groundless injunctions against a host of perfectly safe drug combinations (see my other papers for a detailed analysis of this topic in relation to ST) and various ‘official’ bodies like the WHO and the FDA have been repeatedly guilty of issuing such scientifically groundless injunctions: recent examples are warnings about ‘serotonergic’ drugs precipitating ST if combined with the anti-emetic 5-HT₃ antagonists, and about ST with Triptans. Such ‘cry-wolf’ warnings are time-wasting and confuse and misdirect practicing clinicians. So, adopting the careful conservative approach is not always the ‘win-win’ scenario that such cautious analysts suppose it to be.

In routine clinical practice the maxim ‘start low and go slow’ is wisely adhered to. When a changeover is being considered, or is indicated, in a patient who is *severely* ill, and in danger because of that, a degree of risk, imagined or real, is acceptable. In less urgent circumstances the patient and clinician are usually well-advised to opt for a cautious approach.

In more than 50 years of MAOI use there are only a few reports of supposed difficulties: these are non-specific and not indicative of a cause-effect relationship involving an MAOI-MAOI interaction [365-369]. It is notable that of these incomplete and unconvincing reports, one suggests subarachnoid haemorrhage and another serotonin toxicity. It is highly unlikely that both of these represent a cause-effect interaction, because the required mechanism is quite different for each of them. The most parsimonious explanation is that neither of them represent a cause-effect relationship between the changing drug regime and the outcome reported. As with so many case-reports these ones also contain insufficient information to draw reliable conclusions.

It is well recognised that abrupt cessation of antihypertensive treatment can cause rebound hypertension; indeed, this is a not uncommon presentation in emergency departments. It is forgotten that MAOIs are anti-hypertensive drugs, and, as I have reviewed elsewhere, were indeed used for the treatment of hypertension in the 1960s and 1970s. I have seen patients who have developed high blood pressure when well-meaning primary-care physicians have stopped the ‘dangerous-old-antidepressants-we-don’t-use-anymore’ that the (dodderly old) specialist has been giving because he does not know about the wonderful new drugs we now have (that the young lady drug rep in the short skirt who took me to lunch told me all about).

Furthermore, I have seen patients on long-term antidepressant MAOI treatment who have clearly developed idiopathic hypertension during the course of that treatment, which was ‘disguised’ by the MAOI. These patients then had substantial rises in blood pressure on cessation of the MAOI. Indeed, one of them actually had a small CVA whilst waiting for an appointment to see me, to decide on future treatment. The primary care doctor had already (unilaterally) instructed her to cease the tablets prior to the appointment with me. If that patient had already restarted another MAOI then where would the blame for her CVA have been laid?

The following are the relevant contributions that I am aware of, in the published literature relating to direct MAOI to MAOI change-overs [365-375]: re Torre et al. see

‘case 2’. None of them are very helpful, nor do they provide a substantive basis for prohibition of a direct swap-over of one MAOI to another MAOI. One paper reports a small series of 8 cases where it was accomplished without a problem [376]. I have personally done it without any problem, but only on a couple of occasions, as have associates and various people who have been in contact via my web site.

Opinion: if there is *good reason* to swap rapidly***, do it, because there is; 1) no theoretical basis to suggest it might be contra-indicated, and 2) existing reports do not constitute evidence to the contrary, and 3) it has clearly been done many times without any problem.

***For example, I remember a patient who had failed to improve with a long course of ECT, which had involved a long stay in hospital 1000 km away from home, whom had started phenelzine and was experiencing rapid weight gain and massively swollen the legs, remained suicidal, but dreaded a return to hospital. A direct swap to Parnate was successful and uneventful and led to great improvement.

MAOIs: Miscellaneous observations

NRIs and reducing tyramine sensitivity

Myth: One cannot combine MAOIs with TCAs.

Attenuation of the pressor response to tyramine was demonstrated long ago by the renowned pharmacologist Bernard Brodie. There is a sound pharmacological basis for the proposition that TCAs, by virtue of their NRI potency, attenuate the ‘cheese effect’ [377-382].

I assembled and explained the evidence for this in my TCA review [6] which concluded that those NRIs with a *high* affinity for the NAT (viz. reboxetine, desipramine, oxypropraline, protriptyline and nortriptyline) have all been demonstrated to block the pressor response to tyramine almost completely [379, 383-388], even when it has been potentiated in the presence of MAOIs [389-391]. The early demonstrations of NRI attenuation of the pressor response to tyramine go back a long way, past learning seems to have been lost for a long time [377, 378]. NB Both those last 2 references are from the lab of the famous pharmacologist Bernard Brodie whose early papers are worth studying.

See also update commentary and references [here](#)

This leads to the confident conclusion about an old and bitter-sweet irony: combinations of (non-serotonergic) TCAs or NRIs (e.g. reboxetine etc.) with MAOIs are not risky, they actually make MAOIs safer, not more dangerous, by attenuating, or completely blocking, the pressor response to tyramine, or any other NA releaser (ISA).

If a patient is more than usually tyramine sensitive, they will become less so if a potent NRI like nortriptyline is added to the regime: the greater the dose the greater the attenuation of the pressor response. That is not just a ‘theory’, it is pharmacological fact.

Therapeutic hypotensive effect of MAOIs

Myth: MAOIs cause hypertension and should not be given to hypertensive patients.

MAOIs lower blood pressure: however, the idea that MAOIs raise blood pressure is a widespread misconception, and one of the commoner incorrect statements you will

Check web site for latest V

37

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

see. It is incontrovertibly incorrect: it is only the interaction with releasers like tyramine that produces hypertension.

In the 1960s MAOIs were used to treat hypertension, until better drugs were found [392-394], and indeed using MAOIs for those suffering both depression and hypertension works very well.

The still repeated, but incorrect, prohibition about giving MAOIs to patients with pre-existing hypertension is thus another example of ignorance about pharmacology. Again, the basis on which this opinion has insinuated itself into the literature is impossible to pin down. It just appeared and was unthinkingly repeated by the pharmacologically ignorant presumably because it sounds sensible and responsible. The post 'Parnate-withdrawal' hysteria (1964) was fertile ground for such admonitions. Having been repeated so often it became 'received clinical wisdom', and even 'expert opinion'.

Expert opinion is a capricious beast to be regarded with circumspection.

'Spontaneous' hypertensive episodes

One occasionally sees patients who do get brief episodes of hypertension (but not hypertensive emergencies) — I estimate about 5% of all MAOI takers — lasting a couple of hours, most often following the second dose of the daily regime of MAOI. This seems only to occur with TCP, but not with phenelzine or isocarboxazid. There are only a few reports in the literature [395-398], it may more common than realised, even if only to a minor degree. In my experience this usually gets less over a few weeks. If such elevations are problematically high, symptomatic, or enduring they will be controlled by giving propranolol with the MAOI dose, that also helps reduce exercise-induced postural hypotension by antagonising the noradrenaline beta-receptor-mediated vaso-dilation (in muscle vasculature).

However, there may also be another (presumably very small) group of patients in whom such elevations are related to occult pheochromocytoma, and I have encountered such a case and one or two cases have been reported in the literature [399, 400]. If such elevations are occurring outside of the 1–4 hr. post-dose 'window' then it is suggested that investigations for occult pheochromocytoma, such as a high-resolution scan of the adrenals, should be undertaken to try to rule out a small adrenal tumour. MAOIs will magnify the pressor effect of even a small tumour.

Hypertensive urgencies and emergencies due to tyramine, and the questionable relationship to subarachnoid haemorrhage

Deaths from tyramine/MAOI induced hypertension are *extremely* rare. Indeed, there have been no deaths from MAOI induced hypertension reported in the medical literature for decades (NB I am not suggesting that demonstrates they have not occurred). When De Villiers [401] reviewed this in the UK at the time Parnate was withdrawn in the USA (1964) he noted that of 21,582 treated with MAOIs (between 1960 and 1964), 2% developed headache, 0.27% had the 'hypertensive syndrome'. Fourteen deaths were reported in Great Britain out of an estimated one and a half million patients treated since the introduction of the drugs in 1960. The estimated risk for a patient treated with tranlycypromine was: for headache 2%; for hypertensive crises 0.5% and for death 0.001%. Note that this was when foods had much higher

tyramine levels and *before* tyramine-restricted diets and an understanding of the pressor response.

Furthermore, if one takes into account the reduction of tyramine in all foods these days, and the better dietary advice now available, then the incidence of *serious* hypertensive events is likely to be exceedingly low. Indeed, it is so low that it may be hard to distinguish from the background rate of spontaneously occurring subarachnoid haemorrhage in the general population.

The role of transient episodes of hypertension in the precipitation of subarachnoid haemorrhage has probably been over-estimated. Large numbers of patients are walking around with BPs well in excess of 200 mm/Hg with no (short term) increase in subarachnoid haemorrhage (or cardiac failure) risk: this view is supported by a 2016 paper reviewing 58,000 cases [402]. This shows that the *acute* risk of BPs in the range 180-220 is immeasurably low even in a sample as large as 58,000.

See my recent updated [commentary on this subject](#). In summary, it reviews many physical activities, both indoors and out of doors, which raise blood pressure to 200-250 mmHg, including marathon running and all vigorous sports and weight lifting (BPs are as high as 450 mmHg) [403-407], and the lack of association of any of these BP elevations with significant risk of SAH.

It is important to maintain perspective on the issue of acute BP elevation (which is an unfounded over-concern with many doctors).

The major factor determining a subarachnoid haemorrhage is systemic vulnerability. BPs of 200-250 resulting from many activities, including tyramine ingestion, are only one factor and are somewhat indirectly related to any particular subarachnoid haemorrhage event.

Deaths from MAOI induced hypertension can therefore be inferred to be rarer than fatal reactions to various modern drugs. For example, SSRIs contribute to many deaths from **gastro-intestinal (GI)** bleeding. There is a significantly increased risk of GI bleeding related to SSRIs and the overall mortality rate from GI bleeds is still > 5% [408]. The evidence that SSRIs contribute to increasing the frequency of bleeds is strong [409-415]. So, a GI bleed has a 1/20 risk of death and a serious hypertensive episode more like a 1/1000 risk. It is quite an irony that an SSRI side effect that most doctors do not even know about, or think about, may well cause far more deaths than the feared 'hypertensive crisis'. As they say; *'what the eye does not see, the heart does not grieve over'*.

Medical treatment of hypertension resulting from tyramine ingestion

If excessive tyramine is ingested the blood pressure starts to increase from about half an hour after ingestion (sooner for liquids on an empty stomach), and remains elevated for 1 – 2 hours: the magnitude and duration of that elevation is dose related, so unless a large amount of tyramine has been ingested (50 – 100 mg) the reaction will be short-lived (about one hour).

Current evidence clearly indicates that elevated BP without signs or symptoms of end-organ damage does not require, and should not be given, urgent treatment. This is because hasty and inexpert BP reduction may well do more harm than good (sub-lingual nifedipine is strongly contra-indicated*, see below).

The tyramine reaction is one example of catecholamine-induced hypertensive urgencies, others being the ingestion of amphetamine ('ice'), clonidine withdrawal, and phaeochromocytoma. They probably are comparable.

An SBP of 180 mmHg or more, sustained over 3 measurements in 10 minutes or so, performed in a calm setting with an accurate sphygmomanometer is now referred to as a 'hypertensive urgency'. Only if 'end organ' dysfunction is present it is called a 'hypertensive emergency'. End organ dysfunction is uncommon unless DBP is greater than 130 mmHg [416].

In hypertensive urgencies the treatment aim is to reduce BP slowly over 24-48 hrs. Since tyramine reactions are self-limiting over 2-4 hrs., or rather less with present, typically smaller, tyramine ingestions, it is clear they will *very rarely* require intervention. The exception to this might be when a large (> 100 mg) 'deliberate' tyramine ingestion has occurred and SBP is in excess of 220/130 mm/Hg for a prolonged period (there is then a risk of hypertensive encephalopathy, rather than SAH).

Rapid reduction of hypertension (i.e. within 2-4 hours) carries a serious risk of catastrophic adverse effects [416-419] and such treatment is inadvisable, even if initiated in a specialist hospital setting.

Several of these recent reviews about hypertensive urgencies make very strong statements about premature treatment and about excessively rapid reductions of blood pressure.

Flanigan: "Often the urgency is more in the mind of the treating physician than in the body of the patient ... The compulsive need to treat reaches the pathological in some physicians, especially during the early years in their careers".

Marik: "Rapid reduction of BP may be associated with significant morbidity ... causing ischemia and infarction. It must be lowered in a slow and controlled fashion [over 24 – 48 hrs.] to prevent organ hypoperfusion."

*Sub-lingual nifedipine is very strongly contra-indicated [419-422]. It can result in uncontrollable hypotension and hypo-perfusion which may cause stroke or sudden permanent blindness. Indeed some experts have suggested instant/rapid-release formulations of nifedipine should be prohibited [423, 424] and that it should never be given to patients to self-administer.

Pain and anxiety exacerbate hypertension, so remaining calm and using a benzodiazepine, which will lower BP safely and to a significant and sufficient extent [425-428], is probably the most useful and safe *initial* step. As the above text suggests it is most unlikely that urgent hospital and specialist assessment will be required, unless a very large ingestion of tyramine is suspected (nowadays that would almost have to be 'deliberate'), and observation and BP monitoring shows BP increasing beyond 220 mm/Hg or so over a prolonged time (4 hours), or end organ damage.

A great majority of BP elevations nowadays are going to be mild, from relatively small amounts of tyramine, and will last only an hour or so and require no intervention.

Ceasing Treatment

When MAOIs are ceased, precautions about diet and possible interacting drugs are advisable for four weeks after cessation, especially when starting SRI-type drugs. When SRI-type drugs are being ceased to start MAOIs, five half-lives of the relevant drug ideally should be allowed (various safe bridging strategies are available (see above), but a period of 3 half-lives is often sufficient if the MAOI is being started at a low dose, as is usually advisable.

Acknowledgements

I acknowledge the expertise and assistance of my wife Isobel, who maintains my computers and software, among many other things.

I also acknowledge the companionship and calming influence of more than one generation of fine handsome dogs, without whom my blood pressure would have been higher for longer.

I am indebted to the following people who expended substantial time and effort in advising, encouraging, and correcting this, and previous, versions of the monograph:

Professor Ian Whyte Discipline of Clinical Pharmacology, Faculty of Health, University of Newcastle, Newcastle, NSW, Department of Clinical Toxicology and Pharmacology, Calvary Mater Newcastle Hospital

Dr. Glen Baker, PhD, DSc; Neurochemical Research Unit, Department of Psychiatry, Mackenzie Centre, University of Alberta, Edmonton

Professor Anthony FT Brown MB ChB, FRCP, FRCS(Ed), FRCEM, FACEM Senior Staff Specialist, Department of Emergency Medicine Royal Brisbane and Women's Hospital Brisbane, Queensland 4029

Thanks are due to various others around the world who have helped in various ways and made comments on this monograph.

In view of the above it is appropriate to consider this monograph as 'peer reviewed'.

Help PsychoTropical

I appreciate any help with the costs involved in maintaining PsychoTropical. You can acknowledge the value of this information, and other information on the site, by making a [donation via Paypal at my web site](#).

References

1. White, K. and G.M. Simpson, *Combined MAOI-tricyclic antidepressant treatment: A reevaluation*. A report to the American College of Neuropsychopharmacology, 1980. **Oct**: p. 1-54.
2. Gillman, P.K., *Advances pertaining to the pharmacology and interactions of irreversible nonselective monoamine oxidase inhibitors*. *Journal of Clinical Psychopharmacology*, 2011. **31**(1): p. 66-74.
3. Gillman, P.K., *Combining antidepressants: Understanding Drug Interactions is the Sine Qua Non*. *Advances in Psychiatric Treatment*, 2010. **16**: p. 76-78.
4. Gillman, P.K., *CNS toxicity involving methylene blue: the exemplar for understanding and predicting drug interactions that precipitate serotonin toxicity*. *Journal of Psychopharmacology*, 2011. **25**(3): p. 429-3.

5. Gillman, P.K., *Triptans, Serotonin Agonists, and Serotonin Syndrome (Serotonin Toxicity): A Review*. Headache, 2009. **50**(2): p. 264-272.
6. Gillman, P.K., *Tricyclic antidepressant pharmacology and therapeutic drug interactions updated*. British Journal of Pharmacology 2007. **151**(6): p. 737-48.
7. Gillman, P.K., *A review of serotonin toxicity data: implications for the mechanisms of antidepressant drug action*. Biological Psychiatry, 2006. **59**(11): p. 1046-51.
8. Gillman, P.K., *"Much ado about nothing": monoamine oxidase inhibitors, drug interactions, and dietary tyramine*. CNS Spectr, 2017: p. 1-3.
9. Gillman, P.K., *A reassessment of the safety profile of monoamine oxidase inhibitors: elucidating tired old tyramine myths*. J Neural Transm (Vienna), 2018. **125**(11): p. 1707-1717.
10. Gillman, P.K., S. Feinberg, and L. Fochtmann, *Revitalizing monoamine oxidase inhibitors: A call for action*. CNS spectrums, 2019: p. <http://dx.doi.org/10.1017/S1092852919001196>.
11. Gillman, P.K., *Drug interactions and fluoxetine: a commentary from a clinician's perspective*. Ex Op Drug Saf, 2005. **4**: p. 965-969.
12. Adli, M., et al., *Safety of high-intensity treatment with the irreversible monoamine oxidase inhibitor tranylcypromine in patients with treatment-resistant depression*. Pharmacopsychiatry, 2008. **41**(6): p. 252-7.
13. Parker, G., et al., *Assessing the comparative effectiveness of antidepressant therapies: a prospective clinical practice study*. Journal of Clinical Psychiatry, 2001. **62**(2): p. 117-25.
14. Parker, G., et al., *Are the newer antidepressant drugs as effective as established physical treatments? Results from an Australasian clinical panel review*. Australian and New Zealand Journal of Psychiatry, 1999. **33**(6): p. 874-81.
15. Finberg, J. and P. Gillman, *Pharmacology of MAO-B inhibitors and the cheese reaction*, in *International Review of Neurobiology*, M. Youdim and P. Riederer, Editors. 2011, Elsevier Inc. Academic Press.: Burlington. p. 169-190.
16. Bromberger, B. and F. Percival, *Culture Shock: Principles for Successful Wine-and-Cheese Pairing*. World of Fine Wine, 2007. **16**: p. 139-144.
17. Gillman, P.K., *More on Mrs Murphy's beans: or 'do us a fava'*. Journal of Clinical Psychopharmacology, 2010. **30**(2): p. 215-216.
18. Gillman, P.K., *Ti: myths about monoamine oxidase inhibitors perpetuated*. Australian and New Zealand Journal of Psychiatry, 2009. **43**(11): p. 1084-5.
19. Gillman, P.K., *A systematic review of the serotonergic effects of mirtazapine: implications for its dual action status*. Human Psychopharmacology. Clinical and Experimental, 2006. **21**(2): p. 117-25.
20. Gillman, P.K., *Monoamine oxidase inhibitors, opioid analgesics and serotonin toxicity*. British Journal of Anaesthesia, 2005. **95**(4): p. 434-441.

Check web site for latest V

42

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

21. Anon, *EFSA Panel on Biological Hazards (BIOHAZ); Scientific Opinion on Scientific Opinion on risk based control of biogenic amine formation in fermented foods*. *EFSA Journal*, 2011. **9**(10): p. 2393. [93 pp.].
22. Ladero, V., et al., *Toxicological Effects of Dietary Biogenic Amines*. *Current Nutrition & Food Science*, 2010. **6**: p. 145-156.
23. Zaman, M.Z., et al., *A Review: Microbiological, Physicochemical and Health Impact of High Level of Biogenic Amines in Fish Sauce*. *American Journal of Applied Sciences*, 2009. **6**: p. 1199-1211.
24. Al Bulushi, I., et al., *Biogenic amines in fish: roles in intoxication, spoilage, and nitrosamine formation--a review*. *Critical Reviews in Food Science and Nutrition*, 2009. **49**(4): p. 369-77.
25. Shalaby, A.R., *Significance of biogenic amines to food safety and human health*. *Food Research International*, 1996. **29**: p. 675–690.
26. Mohammed, G.I., et al., *A critical overview on the chemistry, clean-up and recent advances in analysis of biogenic amines in foodstuffs*. *TrAC Trends in Analytical Chemistry*, 2016. **78**: p. 84-94.
27. Scavnicar, A., et al., *Determination of Biogenic Amines in Cheese by Ion Chromatography with Tandem Mass Spectrometry Detection*. *J AOAC Int*, 2018: p. doi: 10.5740/jaoacint.16-0006. [Epub ahead of print].
28. Onal, A., S.E. Tekkeli, and C. Onal, *A review of the liquid chromatographic methods for the determination of biogenic amines in foods*. *Food Chem*, 2013. **138**(1): p. 509-15.
29. Palermo, C., et al., *A multiresidual method based on ion-exchange chromatography with conductivity detection for the determination of biogenic amines in food and beverages*. *Anal Bioanal Chem*, 2013. **405**(2-3): p. 1015-23.
30. Loizzo, M.R., et al., *Technological aspects and analytical determination of biogenic amines in cheese*. *Trends in Food Science & Technology*, 2013. **30**: p. 38-55.
31. Mayer, H.K., G. Fiechter, and E. Fischer, *A new ultra-pressure liquid chromatography method for the determination of biogenic amines in cheese*. *Journal of Chromatography A*, 2010. **1217**: p. 3251–3257.
32. Til, H.P., et al., *Acute and subacute toxicity of tyramine, spermidine, spermine, putrescine and cadaverine in rats*. *Food Chemistry and Toxicology*, 1997. **35**(3-4): p. 337-48.
33. Hungerford, J.M., *Scombroid poisoning: a review*. *Toxicon*, 2010. **56**(2): p. 231-43.
34. Linares, D.M., et al., *Comparative analysis of the in vitro cytotoxicity of the dietary biogenic amines tyramine and histamine*. *Food Chem*, 2016. **197**(Pt A): p. 658-63.
35. Lader, M.H., G. Sakalis, and M. Tansella, *Interactions between sympathomimetic amines and a new monoamine oxidase inhibitor*. *Psychopharmacologia*, 1970. **18**(1): p. 118-23.
36. Bieck, P.R. and K.H. Antonin, *Oral tyramine pressor test and the safety of monoamine oxidase inhibitor drugs: comparison of brofaromine and tranlycypromine in healthy subjects*. *Journal of clinical psychopharmacology*, 1988. **8**(4): p. 237-45.

Check web site for latest V

43

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

37. Korn, A., et al., *Moclobemide, a new reversible MAO inhibitor--interaction with tyramine and tricyclic antidepressants in healthy volunteers and depressive patients*. *Psychopharmacology (Berl)*, 1986. **88**(2): p. 153-7.
38. Bar-Am, O., et al., *Cardiovascular baroreceptor activity and selective inhibition of monoamine oxidase*. *Eur J Pharmacol*, 2012. **15**(683): p. 226-30.
39. European Community. (2011). *BIAMFOOD: Controlling Biogenic Amines in Traditional Food Fermentations*. 2011: p. http://cordis.europa.eu/project/rcn/88095_en.html.
40. Latorre-Moratalla, M.L., et al., *Distribution of aminogenic activity among potential autochthonous starter cultures for dry fermented sausages*. *J Food Prot*, 2010. **73**(3): p. 524-8.
41. Chong, C.Y., et al., *The effects of food processing on biogenic amines formation*. *International Food Research Journal*, 2011. **18**: p. 867-876.
42. Blackwell, B., *Hypertensive Crisis Due to Monoamine-Oxidase Inhibitors*. *Lancet*, 1963. **38**: p. 849-50.
43. McCormick, W.O. and B. Blackwell, *Tranycypromine*. *Lancet*, 1963. **281**(7273): p. 167-168.
44. Blackwell, B., *Toxic Effects of Monoamine-Oxidase Inhibitors*. *Lancet*, 1964. **2**(7351): p. 150-1.
45. Blackwell, B. and E. Marley, *Interaction between Cheese and Monoamine-Oxidase Inhibitors in Rats and Cats*. *Lancet*, 1964. **1**(7332): p. 530-1.
46. Blackwell, B., E. Marley, and A. Ryle, *Hypertensive Crisis Associated with Monoamine-Oxidase Inhibitors*. *Lancet*, 1964. **1**(7335): p. 722-3.
47. Blackwell, B. and L.A. Mabbitt, *Tyramine in Cheese Related to Hypertensive Crises after Monoamine-Oxidase Inhibition*. *Lancet*, 1965. **1**(7392): p. 938-40.
48. Blackwell, B. and E. Marley, *Interactions of yeast extracts and their constituents with monoamine oxidase inhibitors*. *British Journal of Pharmacology*, 1965. **26**(1): p. 142-161.
49. Blackwell, B., E. Marley, and L.A. Mabbitt, *Effects of Yeast Extract after Monoamine-Oxidase Inhibition*. *Lancet*, 1965. **1**(7392): p. 940-3.
50. Blackwell, B., et al., *Hypertensive interactions between monoamine oxidase inhibitors and foodstuffs*. *Br J Psychiatry*, 1967. **113**(497): p. 349-65.
51. Blackwell, B., L. Mabbitt, and E. Marley, *Histamine and tyramine content of yeast products*. *Journal of Food Science*, 1969. **34**(1): p. 47-51.
52. Kosikowski, K.V., *A quantitative appraisal of the free amino acids in foreign type cheese*. *Journal of Dairy Science*, 1954. **37**(2): p. 167-172.
53. Bullock, D. and O. Irvine, *A chromatographic study of Cheddar cheese ripening*. *Journal of Dairy Science*, 1956. **39**(9): p. 1229-1235.
54. Flockhart, D.A., *Dietary restrictions and drug interactions with monoamine oxidase inhibitors: an update*. *J Clin Psychiatry*, 2012. **73 Suppl 1**: p. 17-24.

Check web site for latest V

44

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

55. Zajecka, J.M. and A.M. Zajecka, *A clinical overview of monoamine oxidase inhibitors: pharmacological profile, efficacy, safety/tolerability, and strategies for successful outcomes in the management of major depressive disorders*. *Psych Annals*, 2014. **44**(11): p. 513-523.
56. Wimbiscus, M., O. Kostenko, and D. Malone, *MAO inhibitors: risks, benefits, and lore*. *Cleve Clin J Med*, 2010. **77**(12): p. 859-82.
57. O'Brien, V., *The Monoamine Oxidase Inhibitors: Relics Reconsidered*. *Psychiatric Annals*, 2011. **41**(3): p. 176-183.
58. Rapaport, M.H., *Dietary restrictions and drug interactions with monoamine oxidase inhibitors: the state of the art*. *J Clin Psychiatry*, 2007. **68 Suppl 8**: p. 42-6.
59. Fiechter, G., G. Sivec, and H.K. Mayer, *Application of UHPLC for the simultaneous analysis of free amino acids and biogenic amines in ripened acid-curd cheeses*. *J Chromatogr B*, 2013. **927**: p. 191-200.
60. Bunkova, L., et al., *The effect of ripening and storage conditions on the distribution of tyramine, putrescine and cadaverine in Edam-cheese*. *Food Microbiol*, 2010. **27**(7): p. 880-8.
61. Spizzirri, G.U., et al., *Determination of biogenic amines in different cheese samples by LC with evaporative light scattering detector*. *Journal of Food Composition and Analysis*, 2013. **29**: p. 43–51.
62. Linares, D.M., et al., *Factors Influencing Biogenic Amines Accumulation in Dairy Products*. *Front Microbiol*, 2012. **3**: p. 180.
63. Ibrahim, E.M. and A.A. Amer, *Comparison of biogenic amines levels in different processed cheese varieties with regulatory specifications*. *World Journal of Dairy and Food Science*, 2010. **5**: p. 127-133.
64. Schirone, M., et al., *Biogenic amines in italian pecorino cheese*. *Front Microbiol*, 2012. **3**: p. 171.
65. Rea, M.S., et al., *Development of enterococci and production of tyramine during the manufacture and ripening of Cheddar cheese*. *Irish Journal of Agricultural and Food Research*, 2004. **43**: p. 247–258.
66. Bonczar, G., et al., *The range of protein hydrolysis and biogenic amines content in selected acid-and rennet-curd cheeses*. *Chemical Papers*, 2018. **72**(10): p. 2599-2606.
67. De Vuyst, A., W. Vervack, and M. Foulon, *Détection d'amines non volatiles dans quelques fromages*. *Le Lait*, 1976. **557**: p. 414-422.
68. Komprda, T., et al., *Some factors influencing biogenic amines and polyamines content in Dutch-type semi-hard cheese*. *Eur Food Res Technol*, 2008. **227**: p. 29–36.
69. Komprda, T., et al., *Tyramine production in Dutch-type semi-hard cheese from two different producers*. *Food Microbiol*, 2008. **25**(2): p. 219-27.
70. Horwitz, D., et al., *Monoamine Oxidase Inhibitors, Tyramine, and Cheese*. *JAMA*, 1964. **188**: p. 1108-10.

71. Asatoor, A.M., A.J. Levi, and M.D. Milne, *Tranylcypromine and Cheese*. *Lancet*, 1963. **2**(7310): p. 733-4.
72. Colonna, P. and J. Adda, *Dosage de la Tyramine, Tryptamine et Histamine dans Quelques Fromages Français*. *Lait*, 1976. **56**: p. 143-153.
73. Voight, M. and R. Eitenmiller, *Fluorescent quantitation of biologically active amines in foods with 7 chloro 4 nitro benzo-furazan*. *Journal of Food Science*, 1974. **39**.
74. Samkova, E., E. Dadáková, and T. Pelikánová, *Changes in biogenic amine and polyamine contents in smear-ripened cheeses during storage*. *European Food Research and Technology*, 2013. **237**(3): p. 309-314.
75. Coton, M., et al., *Diversity and assessment of potential risk factors of Gram-negative isolates associated with French cheeses*. *Food microbiology*, 2012. **29**(1): p. 88-98.
76. Valsamaki, K., A. Michaelidou, and A. Polychroniadou, *Biogenic amine production in Feta cheese*. *Food chemistry*, 2000. **71**(2): p. 259-266.
77. Komprda, T., V. Dohnal, and R. Závodníková, *Contents of Some Biologically Active Amines in a Czech Blue-vein Cheese*. *Czech J Food Sci*, 2008. **26**: p. 428–440.
78. Novella-Rodriguez, S., et al., *Distribution of biogenic amines and polyamines in cheese*. *Journal of Food Science*, 2003. **68**: p. 750-755.
79. Novella-Rodriguez, S., M.T. Veciana-Nogues, and M.C. Vidal-Carou, *Biogenic amines and polyamines in milks and cheeses by iron high performance liquid chromatography*. *Journal of Agricultural and Food Chemistry*, 2000. **48**: p. 5117–5123.
80. Novella-Rodriguez, S., et al., *Influence of starter and nonstarter on the formation of biogenic amine in goat cheese during ripening*. *J Dairy Sci*, 2002. **85**(10): p. 2471-8.
81. Gezginc, Y., et al., *Biogenic amines formation in Streptococcus thermophilus isolated from home-made natural yogurt*. *Food Chem*, 2013. **138**(1): p. 655-62.
82. Cho, T.Y., et al., *Evaluation of biogenic amines in Korean commercial fermented foods*. *Korean Journal of Food Science and Technology*, 2006. **38**: p. 730-737.
83. Li, J., et al., *Safety evaluation in vitro of Enterococcus durans from Tibetan traditional fermented yak milk*. *J Microbiol*, 2011. **49**(5): p. 721-8.
84. Rizzello, C.G., et al., *Effect of sourdough fermentation on stabilisation, and chemical and nutritional characteristics of wheat germ*. *Food Chemistry*, 2010. **119**: p. 1079–1089.
85. Ozdestan, O., E. Alpözen, and G. Güven, *Monitoring of biogenic amines in Kumru: a traditional fermented cereal food*. *International Journal of Food Properties*, 2012. **15**: p. 972-981.
86. Ozdestan, O. and A. Uren, *Biogenic Amine Content of Tarhana: A Traditional Fermented Food*. *International Journal of Food Properties*, 2011. **16**: p. 416-428.
87. Populin, T., et al., *A survey on the presence of free glutamic acid in foodstuffs, with and without added monosodium glutamate*. *Food Chemistry*, 2007. **104**: p. 1712-1717

Check web site for latest V

46

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

88. Shulman, K.I., et al., *Dietary restriction, tyramine, and the use of monoamine oxidase inhibitors*. J Clin Psychopharmacol, 1989. **9**(6): p. 397-402.
89. Toro-Funes, N., et al., *Biologically active amines in fermented and non-fermented commercial soybean products from the Spanish market*. Food Chem, 2015. **173**: p. 1119-24.
90. Shukla, S., J.K. Kim, and M. Kim, *Occurrence of Biogenic Amines in Soybean Food Products*. Soya bean and health, 2011: p. 181-193.
91. Guidi, L.R., M. Beatriz, and A. Gloria, *Bioactive amines in soy sauce: Validation of method, occurrence and potential health effects*. Food Chem, 2012. **133**: p. 323–328.
92. Kim, B., B.Y. Byun, and J.H. Mah, *Biogenic amine formation and bacterial contribution in Natto products*. Food Chem, 2012. **135**(3): p. 2005-11.
93. Ibe, A., et al., *[Production of tyramine in "moromi" mash during soy sauce fermentation]*. Shokuhin Eiseigaku Zasshi, 2003. **44**(5): p. 220-6.
94. Yongmeia, L., et al., *Biogenic amines in Chinese soy sauce*. Food Control, 2009. **20**: p. 593-597.
95. Stute, R.K., et al., *Biogenic amines in fish and soy sauce* European Food Research and Technology, 2002. **215**: p. 101-107.
96. Kung, H.-S., Y. Tsaia, and C.I. Weib, *Histamine and other biogenic amines and histamine-forming bacteria in miso products*. Food Chemistry, 2007. **101**: p. 351-356.
97. Hana, B.-Z., F.M. Romboutsb, and M.J.R. Nout, *Amino acid profiles of sufu, a Chinese fermented soybean food*. Journal of Food Composition and Analysis, 2004. **17**: p. 689-698.
98. Kung, H.-S., et al., *Histamine contents and histamine-forming bacteria in sufu products in Taiwan*. Food Control, 2007. **18**: p. 381-386.
99. Byun, B.Y. and J.H. Mah, *Occurrence of biogenic amines in Miso, Japanese traditional fermented soybean paste*. J Food Sci, 2012. **77**(12): p. T216-23.
100. Tsai, C.H., S.C. Chang, and H.-S. Kung, *Histamine contents and histamine-forming bacteria in natto products in Taiwan*. Food Control, 2007. **18**: p. 1026-1030.
101. Shukla, S., et al., *Determination of biogenic amines in Korean traditional fermented soybean paste (Doenjang)*. Food Chem Toxicol, 2010. **48**(5): p. 1191-5.
102. Saaid, M.B., et al., *Determination of biogenic amines in selected Malaysian food*. Food Chemistry, 2009. **113**: p. 1356-1362.
103. Krausová, P., et al., *Content of biologically active polyamines in livers of cattle, pigs and chickens after animal slaughter*. Meat Science, 2006. **73**: p. 640-644.
104. Hedberg, D.L., M.W. Gordon, and B.C. Blueck, *Six cases of hypertensive crisis in patients on tranlycypromine after eating chicken livers*. American Journal of Psychiatry, 1966. **122**: p. 933–937.
105. Boulton, A.A., B. Cookson, and R. Paulton, *Hypertensive crisis in a patient on MAOI antidepressants following a meal of beef liver*. Can Med Assoc J, 1970. **102**(13): p. 1394-5.

Check web site for latest V

47

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

106. Galgano, F., et al., *Role of biogenic amines as index of freshness in beef meat packed with different biopolymeric materials*. Food Research International, 2009. **42**: p. 1147-1152.
107. Krausová, P., et al., *Changes in the content of biologically active polyamines during storage and cooking of pig liver*. Meat Science, 2007. **77**: p. 269–274.
108. Kalac, P. and M.B.A. Glória, *Biogenic amines in cheeses, wines, beers and sauerkraut, in Biological Aspects of Biogenic Amines, Polyamines and Conjugates*, G. Dandriofosse, Editor. 2009, Transworld Research Network: Kerala, India. p. 267-309.
109. Kalac, P., *Biologically active polyamines in beef, pork and meat products: A review* Meat Science, 2006. **73**: p. 1-11.
110. Kalac, P., et al., *Contents of polyamines in selected foods*. Food Chemistry, 2005. **90**: p. 561–564.
111. Jairath, G., et al., *Biogenic amines in meat and meat products and its public health significance: a review*. Journal of Food Science and Technology, 2015. **52**: p. 6835-6846.
112. Vinci, G. and M.L. Antonelli, *Biogenic amines: quality index of freshness in red and white meat* Food Control, 2002. **13**: p. 519-524
113. Ruiz-Capillas, C. and F. Jimenez-Colmenero, *Biogenic amines in meat and meat products*. Crit Rev Food Sci Nutr, 2004. **44**(7-8): p. 489-99.
114. Kozová, M., P. Kalač, and T. Pelikánová, *Changes in the content of biologically active polyamines during beef loin storage and cooking*. Meat Science, 2009. **81**: p. 607-611.
115. Nadon, C.A., M.A. Ismond, and R. Holley, *Biogenic amines in vacuum-packaged and carbon dioxide-controlled atmosphere-packaged fresh pork stored at -1.50 degrees C*. J Food Prot, 2001. **64**(2): p. 220-7.
116. Kozova, M., P. Kalač, and T. Pelikánová, *Contents of biologically active polyamines in chicken meat, liver, heart and skin after slaughter and their changes during meat storage and cooking*. Food Chemistry, 2009. **116**: p. 419–425.
117. Baston, O., et al., *Refrigerated chicken meat freshness. Correlation between easily hydrolysable nitrogen, ph value and biogenic amine contents*. The Annals of the University Dunarea de Jos of Galati, 2008. **Fascicle VI, II**: p. 37-43.
118. Moreira, A.P.S., et al., *Effect of Aging on Bioactive Amines, Microbial Flora, Physico-Chemical Characteristics, and Tenderness of Broiler Breast Meat*. Poultry Science, 2008. **87**: p. 1868-1873.
119. Kalac, P., *The roles of dietary polyamines in human health and their occurrence in foods*. In A. K. Haghi (Ed.), *Advances in Food Science and Technology* (pp. 91–112). New York: Nova Sci. Publ., 2010.
120. Silva, C.M.G., M. Beatriz, and A. Glória, *Bioactive amines in chicken breast and thigh after slaughter and during storage at 4±1 °C and in chicken-based meat products*. Food Chemistry, 2002. **78**: p. 241–248.

121. Fraqueza, M.J., C.M. Alfaia, and A.S. Barreto, *Biogenic amine formation in turkey meat under modified atmosphere packaging with extended shelf life: Index of freshness*. *Poult Sci*, 2012. **91**(6): p. 1465-72.
122. Arvanitoyannis, I.S. and A.C. Stratakos, *Application of Modified Atmosphere Packaging and Active/Smart Technologies to Red Meat and Poultry: A Review*. *Food and Bioprocess Technology*, 2012. **5**: p. 1423-1446.
123. Dadakova, E., T. Pelikánová, and P. Kalač, *Contents of biologically active polyamines in duck meat and giblets after slaughter and their changes during meat storage and cooking*. *Food Research International*, 2012. **48**: p. 28–33.
124. Rokkaa, M., et al., *Monitoring of the quality of modified atmosphere packaged broiler chicken cuts stored in different temperature conditions: B. Biogenic amines as quality-indicating metabolites*. *Food Control*, 2004. **15**: p. 601–607.
125. Durlu-Ozkaya, F., K. Ayhan, and N. Vural, *Biogenic amines produced by Enterobacteriaceae isolated from meat products*. *Meat Sci*, 2001. **58**(2): p. 163-6.
126. Valero, B.V., et al., *Biogenic Amines and Polyamines and Total Aerobic Count During Storage of Vacuum-Packaged Porcine Kidney, Liver and Spleen* *Food Science and Technology International*, 2005. **11**: p. 337-344
127. Kozová, M., P. Kalač, and T. Pelikánová, *Biologically active polyamines in pig kidneys and spleen: Content after slaughter and changes during cold storage and cooking*. *Meat Science*, 2008. **79**: p. 326-331.
128. Delgado-Pando, G., et al., *Enriched n-3 PUFA/konjac gel low-fat pork liver pate: lipid oxidation, microbiological properties and biogenic amine formation during chilling storage*. *Meat Sci*, 2012. **92**(4): p. 762-7.
129. Ruiz-Capillas, C. and F. Jimenez-Colmenero, *The effect of an argon-containing packaging atmosphere on the quality of fresh pork sausages kept at 1 °C*. *Food Control*, 2010. **21**: p. 1331–1337.
130. Curiel, J.A., et al., *Production of biogenic amines by lactic acid bacteria and enterobacteria isolated from fresh pork sausages packaged in different atmospheres and kept under refrigeration*. *Meat Sci*, 2011. **88**(3): p. 368-73.
131. Lorenzo, J.M., et al., *Biogenic amine content during the manufacture of dry-cured lacón, a Spanish traditional meat product: Effect of some additives*. *Meat Science*, 2007. **77**: p. 287-293.
132. Ruiz-Capillas, C. and F. Jimenez-Colmenero, *Biogenic amine content in Spanish retail market meat products treated with protective atmosphere and high pressure*. *European Food Research and Technology*, 2004. **218**: p. 237–241.
133. Leggio, A., et al., *Dry fermented sausages of Southern Italy: a comparison of free amino acids and biogenic amines between industrial and homemade products*. *J Food Sci*, 2012. **77**(4): p. S170-5.

134. Latorre-Moratalla, M.L., et al., *Control of biogenic amines in fermented sausages: role of starter cultures*. *Front Microbiol*, 2012. **3**: p. 169 doi: 10.3389/fmicb.2012.00169. eCollection 2012.
135. Latorre-Moratalla, M.L., et al., *Biogenic amines in traditional fermented sausages produced in selected European countries*. *Food Chemistry*, 2008. **107**: p. 912–921.
136. Gardini, F., et al., *Modeling the aminogenic potential of Enterococcus faecalis EF37 in dry fermented sausages through chemical and molecular approaches*. *Appl Environ Microbiol*, 2008. **74**(9): p. 2740-50.
137. Bover-Cid, S., et al., *Relationships between microbial population dynamics and putrescine and cadaverine accumulation during dry fermented sausage ripening*. *J Appl Microbiol*, 2009. **106**(4): p. 1397-407.
138. Suzzi, G. and F. Gardini, *Biogenic amines in dry fermented sausages: a review*. *Int J Food Microbiol*, 2003. **88**(1): p. 41-54.
139. Bover-Cid, S., et al., *Amino acid-decarboxylase activity of bacteria isolated from fermented pork sausages*. *Int J Food Microbiol*, 2001. **66**(3): p. 185-9.
140. Komprda, T., et al., *Effect of starter culture and storage temperature on the content of biogenic amines in dry fermented sausages poliean*. *Meat Science*, 2001. **59**: p. 267– 276.
141. Papavergou, E.J., I.N. Savvaidis, and I.A. Ambrosiadis, *Levels of biogenic amines in retail market fermented meat products*. *Food Chem*, 2012. **135**(4): p. 2750-5.
142. Ferreira, I.M. and O. Pinho, *Biogenic amines in Portuguese traditional foods and wines*. *J Food Prot*, 2006. **69**(9): p. 2293-303.
143. Miguelez-Arrizadua, M.J., et al., *Biogenic amines in Spanish fermented sausages as a function of diameter and artisanal or industrial origin*. *Journal of the Science of Food and Agriculture*, 2006. **86**: p. 549–557.
144. Prester, L., *Biogenic amines in fish, fish products and shellfish: a review*. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*, 2011. **28**(11): p. 1547-60.
145. Visciano, P., et al., *Biogenic amines in raw and processed seafood*. *Front Microbiol*, 2012. **3**: p. 188.
146. Zeng, X., et al., *Changes of biogenic amines in Chinese low-salt fermented fish pieces (Suan yu) inoculated with mixed starter cultures*. *International Journal of Food Science & Technology*, 2013. **48**(4): p. 685–692.
147. Emborg, J., et al., *Microbial spoilage and formation of biogenic amines in fresh and thawed modified atmosphere-packed salmon (Salmo salar) at 2 degrees C*. *J Appl Microbiol*, 2002. **92**(4): p. 790-9.
148. Katikou, P., et al., *Relation of Biogenic Amines' Formation with Microbiological and Sensory Attributes in Lactobacillus-Inoculated Vacuum-Packed Rainbow Trout (Oncorhynchus mykiss) Fillets*. *Journal of Agricultural and Food Chemistry*, 2006. **54**: p. 4277–4283.

149. Chytiri, S., et al., *Relation of biogenic amines with microbial and sensory changes of whole and filleted freshwater rainbow trout (Onchorynchus mykiss) stored on ice*. J Food Prot, 2004. **67**(5): p. 960-5.
150. Ruiz-Capillas, C. and A. Moral, *Formation of biogenic amines in bulk-stored chilled hake (Merluccius merluccius L.) packed under atmospheres*. J Food Prot, 2001. **64**(7): p. 1045-50.
151. Özogul, F. and Y. Özogula, *Changes in Biogenic Amines in Herring Stored under Modified Atmosphere and Vacuum Pack*. Journal of Food Science, 2002. **67**: p. 2497-2501.
152. Özogul, F. and Y. Özogula, *Biogenic amine content and biogenic amine quality indices of sardines (Sardina pilchardus) stored in modified atmosphere packaging and vacuum packaging*. Food Chemistry, 2006. **99**: p. 574-578
153. Emborg, J. and P. Dalgaard, *Modelling the effect of temperature, carbon dioxide, water activity and pH on growth and histamine formation by Morganella psychrotolerans*. Int J Food Microbiol, 2008. **128**(2): p. 226-33.
154. Gingerich, T.M., et al., *Biogenic amine survey and organoleptic changes in fresh, stored, and temperature-abused bluefish (Pomatomus saltatrix)*. J Food Prot, 1999. **62**(9): p. 1033-7.
155. Brillet, A.A., et al., *Effect of inoculation of Carnobacterium divergens V41, a biopreservative strain against Listeria monocytogenes risk, on the microbiological, chemical and sensory quality of cold-smoked salmon*. International Journal of Food Microbiology, 2005. **104**: p. 309-324.
156. Jørgensen, L.V., P. Dalgaard, and H.H. Huss, *Multiple Compound Quality Index for Cold-Smoked Salmon (Salmo salar) Developed by Multivariate Regression of Biogenic Amines and pH*. Journal of Agricultural and Food Chemistry, 2000. **48**: p. 2448–2453.
157. Periago, M.J., et al., *Monitoring volatile and nonvolatile amines in dried and salted roes of tuna (Thunnus thynnus L.) during manufacture and storage*. J Food Prot, 2003. **66**(2): p. 335-40.
158. Veciana-Nogues, M.T., A. Marine-Font, and M.C. Vidal-Carou, *Biogenic Amines in Fresh and Canned Tuna. Effects of Canning on Biogenic Amine Contents*. Journal of Agricultural and Food Chemistry, 1997. **45**: p. 4324-4328.
159. Veciana-Nogues, M.T., M.C. Vidal-Carou, and A. Marine-Font, *Histamine and Tyramine in Preserved and Semi-preserved Fish Products*. Journal of Food Science, 2006. **54**: p. 1653 - 1655.
160. Erkan, N., N. Helle, and O. Ozden, *[The content of biogenic amines in canned fish from the Turkish market]*. Berl Munch Tierarztl Wochenschr, 2001. **114**(7-8): p. 241-5.
161. Tsai, Y.-H. and H.-F. Kunga, *Determination of histamine in canned mackerel implicated in a food borne poisoning* Food Control, 2005. **16**: p. 579-585.
162. Shulman, K.I. and S.E. Walker, *Refining the MAOI diet: tyramine content of pizzas and soy products*. J Clin Psychiatry, 1999. **60**(3): p. 191-3.
163. Moret, S., et al., *A survey on free biogenic amine content of fresh and preserved vegetables*. Food Chemistry, 2005. **89**(3): p. 355–361.

Check web site for latest V

51

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

164. Ly, D., et al., *HPLC Analysis of Serotonin, Tryptamine, Tyramine, and the Hydroxycinnamic Acid Amides of Serotonin and Tyramine in Food Vegetables*. Journal of Medicinal Food, 2008. **11**(2): p. 385-389.
165. Kalac, P., S. Švecová, and P. T., *Levels of biogenic amines in typical vegetable products*. Food Chemistry, 2002. **77**: p. 349–351.
166. Lavizzari, T., et al., *Improved method for the determination of biogenic amines and polyamines in vegetable products by ion-pair high-performance liquid chromatography*. J Chromatogr A, 2006. **1129**(1): p. 67-72.
167. Calixto, J.B., et al., *Contribution of natural products to the discovery of the transient receptor potential (TRP) channels family and their functions*. Pharmacol Ther, 2005. **106**(2): p. 179-208.
168. Cayeux, I. and C. Starkenmann, *Sensory characterization of compounds with a trigeminal effect for taste modulation purposes*. Flavour: From Food to Perception, 2016: p. 192-207.
169. Generali, J.A., et al., *Hypertensive crisis resulting from avocados and a MAO inhibitor*. Annals of Pharmacotherapy, 1981. **15**(11): p. 904-906.
170. Borges, C.V., et al., *Bioactive amines changes during the ripening and thermal processes of bananas and plantains*. Food Chemistry, 2019: p. 125020.
171. Quansah, L., *Molecular Basis of Catecholamine Biosynthesis in Banana Fruit*. MSc Thesis. Hebrew University of Jerusalem, 2009.
172. Riggan, R.M., M.J. McCarthy, and P.T. Kissinger, *Identification of salsolinol as a major dopamine metabolite in the banana*. J Agric Food Chem, 1976. **24**(1): p. 189-91.
173. Waalkes, T.P., et al., *Serotonin, norpinephrine, and related compounds in bananas*. Science, 1958. **127**: p. 648-650.
174. Tazoe, M., et al., *Hyperkalemia and hyperdopaminemia induced by an obsessive eating of banana in an anorexia nervosa adolescent*. Brain Dev, 2007. **29**(6): p. 369-72.
175. Martel, C.L., et al., *Transport of DA at the blood-brain barrier of the guinea pig: inhibition by psychotropic drugs and nicotine*. Pharmaceutical Research, 1996. **13**(2): p. 290–295.
176. Guo, L., Y. Zhang, and Q. Li, *Spectrophotometric determination of dopamine hydrochloride in pharmaceutical, banana, urine and serum samples by potassium ferricyanide-Fe(III)*. Anal Sci, 2009. **25**(12): p. 1451-5.
177. Bapat, V.A., et al., *In Vitro Production Of L-dopa In Tissue Cultures Of Banana*. Pharmaceutical Biology, 2000. **38**: p. 271-273.
178. Romphophak, T., et al., *Changes in concentrations of phenolic compounds and polyphenol oxidase activity in banana peel during storage*. Food Preservation Science, 2005. **31**: p. 111-115.
179. Kanazawa, K. and H. Sakakibara, *High Content of Dopamine, a Strong Antioxidant, in Cavendish Banana*. Journal of Agricultural and Food Chemistry, 2000. **48**: p. 844–848.

Check web site for latest V

52

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

180. Ogo, Y., et al., *Banana juice reduces bioavailability of levodopa preparation*. Yakugaku Zasshi, 2005. **125**(12): p. 1009-11.
181. Garfinkel, H.A., *Banana and L-dopa*. Br Med J, 1972. **1**(5795): p. 312.
182. Li, J., et al., *Fermented minced pepper by high pressure processing, high pressure processing with mild temperature and thermal pasteurization*. Innovative Food Science & Emerging Technologies, 2016. **36**: p. 34-41.
183. Zhao, G., et al., *Dopamine transporter inhibitory and antiparkinsonian effect of common flowering quince extract*. Pharmacol Biochem Behav, 2008. **90**(3): p. 363-71.
184. White, J., *Fraud fighter: 'Faked research is endemic in China'* New Scientist, 2012(2891): p. <http://www.newscientist.com/article/mg21628910.300-fraud-fighter-faked-research-is-endemic-in-china.html>.
185. Lavizzari, T., et al., *Occurrence of Biogenic Amines and Polyamines in Spinach and Changes during Storage under Refrigeration*. Journal of Agricultural and Food Chemistry, 2007. **55**: p. 9514–9519.
186. Goyoaga, C., et al., *Content and distribution of vicine, convicine and l -DOPA during germination and seedling growth of two Vicia faba L. varieties*. European Food Research and Technology, 2008. **227**: p. 1537-1542.
187. Ladha, S.S., R. Walker, and H.A. Shill, *Case of neuroleptic malignant-like syndrome precipitated by abrupt fava bean discontinuance*. Mov Disord, 2005. **20**(5): p. 630-1.
188. Katzenschlager, R., et al., *Mucuna pruriens in Parkinson's disease: a double blind clinical and pharmacological study*. J Neurol Neurosurg Psychiatry, 2004. **75**(12): p. 1672-7.
189. Misra, L. and H. Wagner, *Extraction of bioactive principles from Mucuna pruriens seeds*. Indian J Biochem Biophys, 2007. **44**(1): p. 56-60.
190. Tharakan, B., et al., *Anti-Parkinson botanical Mucuna pruriens prevents levodopa induced plasmid and genomic DNA damage*. Phytother Res, 2007. **21**(12): p. 1124-6.
191. Randhir, R. and K. Shetty, *Microwave-induced stimulation of L-DOPA, phenolics and antioxidant activity in fava bean (Vicia faba) for Parkinson's diet* Process Biochemistry, 2003. **39**: p. 1775-1784
192. Hunter, K.R., et al., *Monoamine oxidase inhibitors and L-dopa*. Br Med J, 1970. **3**(5719): p. 388 [1 page].
193. Teychenne, P.F., et al., *Interactions of levodopa with inhibitors of monoamine oxidase and L-aromatic amino acid decarboxylase*. Clin Pharmacol Ther, 1975. **18**(3): p. 273-7.
194. Schildkraut, J.J., et al., *Biochemical and pressor effects of oral D,L-dihydroxy-phenylalanine in patients pretreated with antidepressant drugs*. Ann N Y Acad Sci, 1963. **107**: p. 1005-15.
195. Garcia-Garcia, P., et al., *Biogenic amines in packed table olives and pickles*. J Food Prot, 2001. **64**(3): p. 374-8.

196. Tofalo, R., et al., *Microbiological and chemical profiles of naturally fermented table olives and brines from different Italian cultivars*. *Antonie Van Leeuwenhoek*, 2012. **102**(1): p. 121-31.
197. Kosson, R. and K. Elkner¹, *Effect of Storage Period on Biogenic Amine Content in Sauerkraut* *Vegetable Crops Research Bulletin*, 2010. **73**: p. 151-160.
198. Restuccia, D., et al., *Biogenic Amines as Quality Marker in Organic and Fair-Trade Cocoa-Based Products*. *Sustainability*, 2016. **8**(9): p. 856 doi:10.3390/su8090856.
199. Restuccia, D., et al., *Brewing effect on levels of biogenic amines in different coffee samples as determined by LC-UV*. *Food Chem*, 2015. **175**: p. 143-50.
200. Pastore, P., et al., *Determination of biogenic amines in chocolate by ion chromatographic separation and pulsed integrated amperometric detection with implemented wave-form at Au disposable electrode*. *Journal of Chromatography A*, 2005. **1098**: p. 11-115.
201. Baker, G.B., et al., *Simultaneous extraction and quantitation of several bioactive amines in cheese and chocolate*. *Journal of Chromatography*, 1987. **392**: p. 317-331.
202. Granvogl, M. and P. Schieberle, *Quantification of 3-aminopropionamide in cocoa, coffee and cereal products*. *European Food Research and Technology*, 2007. **225**(5-6): p. 857-863.
203. Mayr, C.M. and P. Schieberle, *Development of stable isotope dilution assays for the simultaneous quantitation of biogenic amines and polyamines in foods by LC-MS/MS*. *J Agric Food Chem*, 2012. **60**(12): p. 3026-32.
204. Baranowska, I. and J. Płonka, *Simultaneous Determination of Biogenic Amines and Methylxanthines in Foodstuff—Sample Preparation with HPLC-DAD-FL Analysis*. *Food Analytical Methods*, 2015. **8**(4): p. 963-972.
205. Spizzirri, U.G., et al., *Application of LC with Evaporative Light Scattering Detector for Biogenic Amines Determination in Fair Trade Cocoa-Based Products*. *Food Analytical Methods*, 2016. **9**(8): p. 2200-2209.
206. Tucker, J., et al., *Unapproved pharmaceutical ingredients included in dietary supplements associated with US Food and Drug Administration warnings*. *JAMA Network Open*, 2018. **1**(6): p. e183337-e183337.
207. Strolin Benedetti, M., K.F. Tipton, and R. Whomsley, *Amine oxidases and monooxygenases in the in vivo metabolism of xenobiotic amines in humans: has the involvement of amine oxidases been neglected?* *Fundam Clin Pharmacol*, 2007. **21**(5): p. 467-80.
208. Bonetta, S., et al., *Detection of biogenic amine producer bacteria in a typical Italian goat cheese*. *Journal of Food Protection*, 2008. **71**(1): p. 205-9.
209. Rabie, M.A., et al., *Biogenic amine contents in selected Egyptian fermented foods as determined by ion-exchange chromatography*. *J Food Prot*, 2011. **74**(4): p. 681-5.
210. Berlin, I., et al., *Determination and comparison of the pressor effect of tyramine during long-term moclobemide and tranlycypromine treatment in healthy volunteers*. *Clin Pharmacol Ther*, 1989. **46**(3): p. 344-51.

211. VanDenBerg, C.M., et al., *Tyramine pharmacokinetics and reduced bioavailability with food*. *J Clin Pharmacol*, 2003. **43**(6): p. 604-9.
212. Ottervanger, J.P., et al., *[Intracranial hemorrhage following use of MAO inhibitor tranylcypromine and beer]*. *Ned Tijdschr Geneesk*, 1993. **137**(18): p. 921-2.
213. Lonvaud-Funel, A., *Biogenic amines in wines: role of lactic acid bacteria*. *FEMS Microbiol Lett*, 2001. **199**(1): p. 9-13.
214. Ancin-Azpilicueta, C., A. González-Marco, and N. AJiménez-Moreno, *Current Knowledge about the Presence of Amines in Wine*. *Critical Reviews in Food Science and Nutrition*, 2008. **48**: p. 257 – 275.
215. Hlabangana, L., S. Hernandez-Cassou, and J. Saurina, *Determination of biogenic amines in wines by ion-pair liquid chromatography and post-column derivatization with 1,2-naphthoquinone-4-sulphonate*. *J Chromatogr A*, 2006. **1130**(1): p. 130-6.
216. Gomez-Alonso, S., I. Hermosin-Gutierrez, and E. Garcia-Romero, *Simultaneous HPLC analysis of biogenic amines, amino acids, and ammonium ion as aminoenone derivatives in wine and beer samples*. *J Agric Food Chem*, 2007. **55**(3): p. 608-13.
217. Jayarajah, C.N., et al., *Analysis of neuroactive amines in fermented beverages using a portable microchip capillary electrophoresis system*. *Annals of Chemistry*, 2007. **79**(21): p. 8162-9.
218. Konakovsky, V., et al., *Levels of histamine and other biogenic amines in high-quality red wines*. *Food Addit Contam Part A Chem Anal Control Expo Risk Assess*, 2011. **28**(4): p. 408-16.
219. Arrieta, M.P. and M.S. Prats-Moya, *Free amino acids and biogenic amines in Alicante Monastrell wines*. *Food Chem*, 2012. **135**(3): p. 1511-9.
220. Ginterova, P., et al., *Determination of selected biogenic amines in red wines by automated on-line combination of capillary isotachopheresis-capillary zone electrophoresis*. *J Chromatogr B Analyt Technol Biomed Life Sci*, 2012. **904**: p. 135-9. doi: 10.1016/j.jchromb.2012.07.018.
221. Henriquez-Aedo, K., et al., *Evaluation of biogenic amines content in Chilean reserve varietal wines*. *Food Chem Toxicol*, 2012. **50**(8): p. 2742-50.
222. Costantini, A., et al., *An Overview on Biogenic Amines in Wine*. *Beverages*, 2019. **5**(1): p. 19.
223. Moreno-Arribas, M.V. and M.C. Polo, *Occurrence of lactic acid bacteria and biogenic amines in biologically aged wines*. *Food Microbiol*, 2008. **25**(7): p. 875-81.
224. Mafra, I., et al., *Evaluation of Biogenic Amines in Some Portuguese Quality Wines by HPLC Fluorescence Detection of OPA Derivatives*. *American journal of Enology and Viticulture*, 1999. **50**: p. 128-132.
225. Herbert, P., et al., *Free amino acids and biogenic amines in wines and musts from the Alentejo region: evolution of amines during alcoholic fermentation and relationship with variety, sub-region and vintage*. *Journal of Food Engineering*, 2005. **66**: p. 315–322.

226. Leitao, M., C.A.P. Marques, and M.V.S. Romao, *A survey of biogenic amines in commercial Portuguese wines*. Food Control, 2005. **16**: p. 199-204.
227. Landete, J.M., et al., *Biogenic amines in wines from three Spanish regions*. J Agric Food Chem, 2005. **53**(4): p. 1119-24.
228. Marcobal, A., et al., *Biogenic amine content of red Spanish wines: comparison of a direct ELISA and an HPLC method for the determination of histamine in wines*. Food Research International, 2005. **38**: p. 387–394.
229. Preti, R., S. Vieri, and G. Vinci, *Biogenic amine profiles and antioxidant properties of Italian red wines from different price categories*. Journal of Food Composition and Analysis, 2015.
230. Hannah, P., V. Glover, and M. Sandler, *Tyramine in wine and beer*. Lancet, 1988. **1**(1): p. 879.
231. Kirschbaum, J., M. A. and H. Brückner, *Determination of biogenic amines in fermented beverages and vinegars by pre-column derivatization with para - nitrobenzyloxycarbonyl chloride (PNZ-Cl) and reversed-phase LC Chromatographia*, 1999. **49**: p. 117-124.
232. Kalac, P. and M. Krizek, *A Review of Biogenic Amines and Polyamines in Beer*. Journal of the Institute of Brewing, 2003. **109**: p. 123–128.
233. Kalac, P., V. Hlavatá, and M. Kůřek, *Concentrations of five biogenic amines in Czech beers and factors affecting their formation*. Food Chemistry, 1997. **58**: p. 209–214.
234. Kalac, P., et al., *Biogenic amine formation in bottled beer*. Food Chemistry, 2002. **79**: p. 431–434.
235. Buňka, F., et al., *Content of biogenic amines and polyamines in beers from the Czech Republic*. Journal of the Institute of Brewing, 2012. **118**: p. 213–216.
236. Pradenas, J., et al., *Occurrence of biogenic amines in beers from Chilean market*. Food Control, 2016. **70**: p. 138-144.
237. Tang, T., et al., *Determination of biogenic amines in beer with pre-column derivatization by high performance liquid chromatography*. J Chromatogr B Analyt Technol Biomed Life Sci, 2009. **877**(5-6): p. 507-12.
238. Cortacero-Ramirez, S., et al., *Determination of biogenic amines in beers and brewing-process samples by capillary electrophoresis coupled to laser-induced fluorescence detection*. Food Chemistry, 2007. **100**: p. 383–389.
239. Izquierdo-Pulido, M., M.C. Vidal-Caour, and A. Marin'e-Font, *Determination of biogenic amines in beers and their raw materials by ion-pair liquid chromatography with postcolumn derivatization*. J. AOAC Int, 1993. **76**: p. 1027–1032.
240. Izquierdo-Pulido, M., M.C. Vidal-Caour, and A. Marin'e-Font, *Histamine and tyramine in beers: contents and relationship with other analytical data*. J. Food Compos. Anal, 1989. **2**: p. 219–227.

241. Izquierdo-Pulido, M., et al., *Biogenic Amines in European Beers*. Journal of Agricultural and Food Chemistry, 1996. **44**: p. 3159-3163.
242. Anli, R.E., et al., *Biogenic Amine Content of Beers Consumed in Turkey and Influence of Storage Conditions on Biogenic Amine Formation*. Journal of the Institute of Brewing, 2006. **112**: p. 267–274.
243. Tailor, S.A., et al., *Hypertensive episode associated with phenelzine and tap beer--a reanalysis of the role of pressor amines in beer*. J Clin Psychopharmacol, 1994. **14**(1): p. 5-14.
244. Loret, S., P. Deloyer, and G. Dandrifosse, *Levels of biogenic amines as a measure of the quality of the beer fermentation process: Data from Belgian samples*. Food Chemistry, 2005. **89**: p. 519-525.
245. Senanayake, N. and S. Vyravanathan, *Histamine reactions due to ingestion of tuna fish (Thunnus argentivittatus) in patients on anti-tuberculosis therapy*. Toxicol, 1981. **19**: p. 184–185.
246. Uragoda, C.G. and S.C. Lodha, *Histamine intoxication in a tuberculous patient after ingestion of cheese*. Tubercle, 1979. **60**(1): p. 59-61.
247. Uragoda, C.G., *Histamine poisoning in tuberculous patients after ingestion of tuna fish*. Am Rev Respir Dis, 1980. **121**(1): p. 157-9.
248. Aloysius, D.J. and C.G. Uragoda, *Histamine poisoning on ingestion of tuna fish*. J Trop Med Hyg, 1983. **86**(1): p. 13-5.
249. Diao, Y.F., *[Histamine-like reaction in tuberculosis patients eating fish containing much histamine under treatment with isoniazid in 277 cases]*. Zhonghua Jie He He Hu Xi Xi Ji Bing Za Zhi, 1986. **9**(5): p. 267-9, 317-8.
250. Morinaga, S., et al., *Histamine poisoning after ingestion of spoiled raw tuna in a patient taking isoniazid*. Intern Med, 1997. **36**(3): p. 198-200.
251. Miki, M., T. Ishikawa, and H. Okayama, *An outbreak of histamine poisoning after ingestion of the ground saury paste in eight patients taking isoniazid in tuberculous ward*. Internal Medicine, 2005. **44**(11): p. 1133-6.
252. Kahana, L.M. and E. Todd, *Histamine intoxication in a tuberculosis patient on isoniazid*. Canadian Disease Weekly Report, 1981. **7**: p. 79-80.
253. Maintz, L. and N. Novak, *Histamine and histamine intolerance*. Am J Clin Nutr, 2007. **85**(5): p. 1185-96.
254. Jarisch, R., *Histamin-Intoleranz. Histamin und Seekrankheit*. 2nd ed. Thieme, Stuttgart, 2004.
255. Ricci, G., et al., *Tryptase serum level as a possible indicator of scombroid syndrome*. Clin Toxicol (Phila), 2010. **48**(3): p. 203-6.
256. Marley, E. and K.M. Wozniak, *Clinical and experimental aspects of interactions between amine oxidase inhibitors and amine re-uptake inhibitors*. Psychological Medicine, 1983. **13**: p. 735-749.

257. Marley, E. and K.M. Wozniak, *Interactions between non-selective amine oxidase inhibitors (MAOI) and other antidepressants*. Brit J Pharmac, 1983. **78**: p. 20p.
258. Marley, E. and K.M. Wozniak, *Interactions between relatively selective amine oxidase (MAOI) inhibitors and clomipramine*. Brit J Pharmac, 1983. **78**: p. 21p.
259. Marley, E. and K.M. Wozniak, *Interactions of a non-selective monoamine oxidase inhibitor, phenelzine, with inhibitors of 5-hydroxytryptamine, dopamine or noradrenaline re-uptake*. Journal of Psychiatric Research, 1984. **18**: p. 173-189.
260. Marley, E. and K.M. Wozniak, *Interactions of non-selective monoamine oxidase inhibitors, tranylcypromine and nialamide, with inhibitors of 5-hydroxytryptamine, dopamine and noradrenaline re-uptake*. Journal of Psychiatric Research, 1984. **18**: p. 191-203.
261. Marley, E. and K.M. Wozniak, *Interactions between relatively selective monoamine oxidase inhibitors and an inhibitor of 5-hydroxytryptamine re-uptake, clomipramine*. Journal of Psychiatric Research, 1985. **19**: p. 597-608.
262. Horton, R., *The Dawn of McScience*. New York Review of Books, 2004. **51**: p. 7-9.
263. Horton, R., *Memorandum by Richard Horton (PI 108). The pharmaceutical industry and medical journals*. UK Parliament: Select Committee on Health. Minutes of Evidence, 2004: p. <https://publications.parliament.uk/pa/cm200405/cmselect/cmhealth/42/4121604.htm>.
264. Karasu, T.B., et al., *Practice guidelines for the treatment of patients with major depressive disorder (revision)*. American Journal of Psychiatry, 2000. **157**: p. (Suppl) 1–45.
265. Malhi, G.S., et al., *Clinical practice recommendations for depression*. Acta Psychiatr Scand Suppl, 2009. **119 (Suppl. 439)**(439): p. 8-26.
266. Anderson, I.M., et al., *Evidence-based guidelines for treating depressive disorders with antidepressants: a revision of the 2000 British Association for Psychopharmacology guidelines*. J Psychopharmacol, 2008. **22**(4): p. 343-96.
267. Frances, A.J., et al., *The expert consensus guidelines for treating depression in bipolar disorder*. J Clin Psychiatry, 1998. **59**(Suppl 4): p. 73-9.
268. Bauer, M., et al., *World federation of societies of biological psychiatry (WFSBP) Guidelines for biological treatment of unipolar depressive disorders, part 1: Acute and continuation treatment of major depressive disorder*. World J Biol Psychiatry, 2002. **3**(1): p. 5-43.
269. Yatham, L.N., et al., *Canadian network for mood and anxiety treatments (CANMAT) guidelines for the management of patients with bipolar disorder: consensus and controversies*. Bipolar Disord, 2005. **7 Suppl 3**: p. 5-69.
270. Heijnen, W.T., et al., *Efficacy of Tranylcypromine in Bipolar Depression: A Systematic Review*. J Clin Psychopharmacol, 2015. **35**(6): p. 700-5.
271. Grady, M.M. and S.M. Stahl, *Practical guide for prescribing MAOIs: debunking myths and removing barriers*. CNS Spectr, 2012. **17**(1): p. 2-10.
272. Mallinger, A.G., et al., *Revisiting the effectiveness of standard antidepressants in bipolar disorder: are monoamine oxidase inhibitors superior?* Psychopharmacol Bull, 2009. **42**(2): p. 64-74.

Check web site for latest V

58

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

273. Fawcett, J., *Why aren't MAOIs used more often?* J Clin Psychiatry, 2009. **70**(1): p. 139-40.
274. Shulman, K.I., et al., *Current prescription patterns and safety profile of irreversible monoamine oxidase inhibitors: a population-based cohort study of older adults.* J Clin Psychiatry, 2009. **70**(12): p. 1681-6.
275. Kennedy, N. and E.S. Paykel, *Treatment and response in refractory depression: results from a specialist affective disorders service.* J Affect Disord, 2004. **81**(1): p. 49-53.
276. Fiedorowicz, J.G. and K.L. Swartz, *The role of monoamine oxidase inhibitors in current psychiatric practice.* Journal of Psychiatric Practice, 2004. **10**(4): p. 239-48.
277. Nutt, D. and P. Glue, *Monoamine oxidase inhibitors: rehabilitation from recent research?* Br J Psychiatry, 1989. **154**: p. 287-91.
278. Petersen, T., et al., *A survey of prescribing practices in the treatment of depression.* Prog Neuropsychopharmacol Biol Psychiatry, 2002. **26**(1): p. 177-87.
279. Paykel, E.S. and J.L. White, *A European study of views on the use of monoamine oxidase inhibitors.* Br J Psychiatry, 1989. **155 (suppl 6)**: p. 9–17.
280. Krishnan, K.R., *Revisiting monoamine oxidase inhibitors.* J Clin Psychiatry, 2007. **68 Suppl 8**: p. 35-41.
281. Thase, M.E., M.H. Trivedi, and A.J. Rush, *MAOIs in the contemporary treatment of depression.* Neuropsychopharmacology, 1995. **12**(3): p. 185-219.
282. Henkel, V., et al., *Treatment of depression with atypical features: a meta-analytic approach.* Psychiatry Res, 2006. **141**(1): p. 89-101.
283. Nierenberg, A.A., et al., *Course and treatment of atypical depression.* J Clin Psychiatry, 1998. **59 Suppl 18**: p. 5-9.
284. Atchley, D.W., *Reevaluation of Tranylcypromine Sulfate.* JAMA, 1964. **189**: p. 763-4.
285. Gillman, P.K., *Moclobemide and the risk of serotonin toxicity (or serotonin syndrome).* Central Nervous System Drug Reviews, 2004. **10**: p. 83-85.
286. Gillman, P.K., *Extracting value from case reports: lessons from serotonin toxicity.* Anaesthesia, 2006. **61**: p. 419-422.
287. Iversen, L., et al., *Neurochemical profiles of some novel psychoactive substances.* Eur J Pharmacol, 2013. **700**(1-3): p. 147-51.
288. Pilgrim, J.L., et al., *Serotonin toxicity involving MDMA (ecstasy) and moclobemide.* Forensic Science International, 2011.
289. Vuori, E., et al., *Death following ingestion of MDMA (ecstasy) and moclobemide.* Addiction, 2003. **98**(3): p. 365-8.
290. Schifano, F., et al., *Novel psychoactive substances of interest for psychiatry.* World Psychiatry, 2015. **14**(1): p. 15-26.

291. Cottencin, O., B. Rolland, and L. Karila, *New designer drugs (synthetic cannabinoids and synthetic cathinones): review of literature*. *Curr Pharm Des*, 2014. **20**(25): p. 4106-11.
292. Meyer, M.R. and H.H. Maurer, *Metabolism of designer drugs of abuse: an updated review*. *Curr Drug Metab*, 2010. **11**(5): p. 468-82.
293. Musselman, M.E. and J.P. Hampton, "Not for human consumption": a review of emerging designer drugs. *Pharmacotherapy*, 2014. **34**(7): p. 745-57.
294. Coupland, C., et al., *Antidepressant use and risk of suicide and attempted suicide or self harm in people aged 20 to 64: cohort study using a primary care database*. *BMJ*, 2015. **350**: p. h517.
295. Tidemalm, D., et al., *Age-specific suicide mortality following non-fatal self-harm: national cohort study in Sweden*. *Psychol Med*, 2015. **45**(8): p. 1699-707.
296. Nordentoft, M., P.B. Mortensen, and C.B. Pedersen, *Absolute risk of suicide after first hospital contact in mental disorder*. *Arch Gen Psychiatry*, 2011. **68**(10): p. 1058-64.
297. Harris, E.C. and B. Barraclough, *Suicide as an outcome for mental disorders. A meta-analysis*. *British Journal of Psychiatry*, 1997. **170**: p. 205-28.
298. Simon, G.E., et al., *Suicide risk during antidepressant treatment*. *American Journal of Psychiatry*, 2006. **163**(1): p. 41-7.
299. Simon, G.E., et al., *Risk of suicide attempt and suicide death in patients treated for bipolar disorder*. *Bipolar Disord*, 2007. **9**(5): p. 526-30.
300. Philippe, A., [Suicide: epidemiological data]. *Rev Prat*, 2011. **61**(2): p. 175-9, 182-3.
301. Olin, B., et al., *Mortality and suicide risk in treatment-resistant depression: an observational study of the long-term impact of intervention*. *PLoS One*, 2012. **7**(10): p. e48002.
302. Pompili, M., et al., *Epidemiology of suicide in bipolar disorders: a systematic review of the literature*. *Bipolar Disord*, 2013. **15**(5): p. 457-90.
303. Crump, C., et al., *Comorbidities and mortality in bipolar disorder: a Swedish national cohort study*. *JAMA Psychiatry*, 2013. **70**(9): p. 931-9.
304. Blair-West, G.W., et al., *Lifetime suicide risk in major depression: sex and age determinants*. *J Affect Disord*, 1999. **55**(2-3): p. 171-8.
305. Angst, J., et al., *Suicide in 406 mood-disorder patients with and without long-term medication: a 40 to 44 years' follow-up*. *Arch Suicide Res*, 2005. **9**(3): p. 279-300.
306. Shorter, E., *Before prozac: the troubled history of mood disorders in psychiatry*. 2009: Oxford University Press.
307. Isbister, G.K., et al., *Moclobemide poisoning: toxicokinetics and occurrence of serotonin toxicity*. *British Journal of Clinical Pharmacology*, 2003. **56**: p. 441-450.
308. Isbister, G.K., P. McGettigan, and A. Dawson, *A fatal case of moclobemide-citalopram intoxication*. *Journal of Analytical Toxicology*, 2001. **25**(8): p. 716-7.

Check web site for latest V

60

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

309. Buckett, W.R., P.C. Thomas, and G.P. Luscombe, *The pharmacology of sibutramine hydrochloride (BTS 54 524), a new antidepressant which induces rapid noradrenergic down-regulation*. Prog Neuropsychopharmacol Biol Psychiatry, 1988. **12**(5): p. 575-84.
310. King, D.J. and N. Devaney, *Clinical pharmacology of sibutramine hydrochloride (BTS 54524), a new antidepressant, in healthy volunteers*. British Journal of Clinical Pharmacology, 1988. **26**(5): p. 607-11.
311. Dujovne, C.A., et al., *Effects of sibutramine on body weight and serum lipids: a double-blind, randomized, placebo-controlled study in 322 overweight and obese patients with dyslipidemia*. Am Heart J, 2001. **142**(3): p. 489-97.
312. Heshka, S., et al., *Anorectic activity and safety of sibutramine 10 mg per day in obese subjects*. North American Association for the Study of Obesity, Milwaukee., 1993.
313. Siramshetty, V.B., et al., *WITHDRAWN-a resource for withdrawn and discontinued drugs*. Nucleic Acids Res, 2015.
314. Young, C.S., R. Mason, and S.J. Hill, *Inhibition by H1-antihistamines of the uptake of noradrenaline and 5-HT into rat brain synaptosomes*. Biochem Pharmacol, 1988. **37**(5): p. 976-8.
315. Yeh, S.Y., et al., *Effects of antihistamines on 3, 4-methylenedioxymethamphetamine-induced depletion of serotonin in rats*. Synapse, 1999. **33**(3): p. 207-17.
316. Lidbrink, P., G. Jonsson, and K. Fuxe, *The effect of imipramine-like drugs and antihistamine drugs on uptake mechanisms in the central noradrenaline and 5-hydroxytryptamine neurons*. Neuropharmacology, 1971. **10**(5): p. 521-536.
317. Carlsson, A., J. Jonason, and M. Lindqvist, *Demonstration of extra neuronal 5-Hydroxytryptamine accumulation in brain following membrane-pump blockade by chlorimipramine*. Brain Research, 1969. **12**: p. 456-460.
318. Carlsson, A. and M. Lindqvist, *Central and peripheral monoaminergic membrane-pump blockade by some addictive analgesics and antihistamines*. Journal of Pharmacy Pharmacology and Chemotherapy, 1969. **21**: p. 460-464.
319. Miyata, S., et al., *Chlorpheniramine exerts anxiolytic-like effects and activates prefrontal 5-HT systems in mice*. Psychopharmacology (Berl), 2011. **213**(2-3): p. 441-52.
320. Gruetter, C.A., et al., *Potentialiation of 5-hydroxytryptamine-induced contraction in rat aorta by chlorpheniramine, citalopram and fluoxetine*. European Journal of Pharmacology, 1992. **217**: p. 109-118.
321. Monte, A.A., R. Chuang, and M. Bodmer, *Dextromethorphan, chlorphenamine and serotonin toxicity: case report and systematic literature review*. British Journal of Clinical Pharmacology, 2010. **70**(6): p. 794-8.
322. Sakurai, E., et al., *Effects of d- and dl-chlorpheniramine on serotonin and 5-hydroxyindoleacetic acid levels in the regional parts of rat brain*. Yakubutsu Seishin Kodo, 1991. **11**(4): p. 237-44.
323. Schwartz, A.R., A.F. Pizon, and D.E. Brooks, *Dextromethorphan-induced serotonin syndrome*. Clin Toxicol (Phila), 2008: p. 1-3.

Check web site for latest V

61

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

324. Shishido, S., R. Oishi, and K. Saeki, *In vivo effects of some histamine H1-receptor antagonists on monoamine metabolism in the mouse brain*. Naunyn-Schmiedeberg's Archives of Pharmacology, 1991. **343**(2): p. 185-189.
325. Sitte, H.H. and M. Freissmuth, *Amphetamines, new psychoactive drugs and the monoamine transporter cycle*. Trends Pharmacol Sci, 2015. **36**(1): p. 41-50.
326. Heal, D.J., et al., *Amphetamine, past and present--a pharmacological and clinical perspective*. J Psychopharmacol, 2013. **27**(6): p. 479-96.
327. Israel, J.A., *Combining Stimulants and Monoamine Oxidase Inhibitors: A Reexamination of the Literature and a Report of a New Treatment Combination*. Prim Care Companion CNS Disord, 2015. **17**(6): p. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4805402/>.
328. Jackson, H., H. Rowley, and D. Hackett, *Comparison of the effects of equivalent doses of lisdexamfetamine dimesylate and d-amphetamine on extracellular concentrations of striatal dopamine, locomotor activity and plasma amphetamine concentrations in freely moving rats*. 2011: p. <http://www.SfN.org> (accessed August 2012).
<http://www.abstractsonline.com/plan/ViewAbstract.aspx?cKey=e73b3b2a-b901-436f-8f5a-dbef5de2ac4c&mID=2773&mKey=%7b8334BE29-8911-4991-8C31-32B32DD5E6C8%7d&sKey=2c0c2336-9990-4c18-b809-bafc7054fefa>.
329. Krishnan, S. and S. Moncrief, *An evaluation of the cytochrome p450 inhibition potential of lisdexamfetamine in human liver microsomes*. Drug Metab Dispos, 2007. **35**(1): p. 180-4.
330. Ermer, J., M. Corcoran, and P. Martin, *Lisdexamfetamine Dimesylate Effects on the Pharmacokinetics of Cytochrome P450 Substrates in Healthy Adults in an Open-Label, Randomized, Crossover Study*. Drugs R D, 2015. **15**(2): p. 175-85.
331. Ermer, J.C., B.A. Adeyi, and M.L. Pucci, *Pharmacokinetic variability of long-acting stimulants in the treatment of children and adults with attention-deficit hyperactivity disorder*. CNS Drugs, 2010. **24**(12): p. 1009-25.
332. Ermer, J.C., M. Pennick, and G. Frick, *Lisdexamfetamine Dimesylate: Prodrug Delivery, Amphetamine Exposure and Duration of Efficacy*. Clin Drug Investig, 2016. **36**(5): p. 341-56.
333. Pennick, M., *Absorption of lisdexamfetamine dimesylate and its enzymatic conversion to d-amphetamine*. Neuropsychiatr Dis Treat, 2010. **6**: p. 317-27.
334. Ermer, J.C., et al., *Pharmacokinetics of lisdexamfetamine dimesylate after targeted gastrointestinal release or oral administration in healthy adults*. Drug Metab Dispos, 2012. **40**(2): p. 290-7.
335. Rowley, H.L., et al., *Lisdexamfetamine and immediate release d-amphetamine - differences in pharmacokinetic/pharmacodynamic relationships revealed by striatal microdialysis in freely-moving rats with simultaneous determination of plasma drug concentrations and locomotor activity*. Neuropharmacology, 2012. **63**(6): p. 1064-74.
336. Hutson, P.H., M. Pennick, and R. Secker, *Preclinical pharmacokinetics, pharmacology and toxicology of lisdexamfetamine: a novel d-amphetamine pro-drug*. Neuropharmacology, 2014. **87**: p. 41-50.

Check web site for latest V

62

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropical@gmail.com

337. Rothman, R.B. and M.H. Baumann, *Serotonin releasing agents. Neurochemical, therapeutic and adverse effects*. Pharmacol Biochem Behav, 2002. **71**(4): p. 825-36.
338. Rothman, R.B., et al., *Amphetamine-type central nervous system stimulants release norepinephrine more potently than they release dopamine and serotonin*. Synapse, 2001. **39**(1): p. 32-41.
339. Rothman, R.B., et al., *In vitro characterization of ephedrine-related stereoisomers at biogenic amine transporters and the receptorome reveals selective actions as norepinephrine transporter substrates*. J Pharmacol Exp Ther, 2003. **307**(1): p. 138-45.
340. Fenwick, M.J. and C.L. Muwanga, *Anaphylaxis and monoamine oxidase inhibitors--the use of adrenaline*. Journal of Accident and Emergency Medicine, 2000. **17**(2): p. 143-4.
341. Griesemer, E., et al., *Potentiating effect of iproniazid on the pharmacological action of sympathomimetic amines*. Experimental Biology and Medicine, 1953. **84**(3): p. 699-701.
342. Burn, J.H., F.J. Philpot, and U. Trendelenburg, *Effect of denervation on enzymes in iris and blood vessels*. British Journal of Pharmacology, 1954. **9**: p. 423-428.
343. Corne, S. and J. Graham, *The effect of inhibition of amine oxidase in vivo on administered adrenaline, noradrenaline, tyramine and serotonin*. The Journal of physiology, 1957. **135**(2): p. 339-349.
344. Markowitz, J.S., S.D. Morrison, and C.L. DeVane, *Drug interactions with psychostimulants*. International Clinical Psychopharmacology, 1999. **14**(1): p. 1-18.
345. Feinberg, S.S., *Combining stimulants with monoamine oxidase inhibitors: a review of uses and one possible additional indication*. J Clin Psychiatry, 2004. **65**(11): p. 1520-4.
346. Rivers, N. and B. Horner, *Possible lethal interaction between Nardil and dextromethorphan*. Canadian Medical Association Journal, 1970. **103**([letter]): p. 85.
347. Shamsie, S.J. and C. Barriga, *The hazards of monoamine oxidase inhibitors in disturbed adolescents*. Canadian Medical Association Journal, 1971. **104**([letter]): p. 715.
348. Asch, D.A. and R.M. Parker, *The Libby Zion case: One step forward or two steps backward?* New England Journal of Medicine, 1988. **318**: p. 771-775.
349. Kaplan, R.L., *The Libby Zion case*. Annals of Internal Medicine, 1991. **115**(12 (letter)): p. 985.
350. Gillman, P.K., *Serotonin Syndrome: History and Risk*. Fundamental and Clinical Pharmacology, 1998. **12**(5): p. 482-491.
351. Rim, C.L. and M.J. Gitlin, *Ziprasidone, monoamine oxidase inhibitors, and the serotonin syndrome*. Journal of Clinical Psychopharmacology, 2010. **30**(4): p. 470-1.
352. Sun-Edelstein, C., S.J. Tepper, and R.E. Shapiro, *Drug-induced serotonin syndrome: a review*. Expert Opinion on Drug Safety, 2008. **7**(5): p. 587-96.
353. Evans, R.W., *The FDA alert on serotonin syndrome with combined use of SSRIs or SNRIs and Triptans: an analysis of the 29 case reports*. MedGenMed, 2007. **9**(3): p. 48.

Check web site for latest V

63

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

354. Evans, R.W., *Concomitant triptan and SSRI or SNRI use: what is the risk for serotonin syndrome?* Headache, 2008. **48**(4): p. 639-40.
355. Tepper, S.J., *Serotonin syndrome: SSRIs, SNRIs, triptans, and current clinical practice.* Headache, 2012. **52**(2): p. 195-7.
356. el-Ganzouri, A.R., et al., *Monoamine oxidase inhibitors: should they be discontinued preoperatively?* Anesthesia and Analgesia, 1985. **64**(6): p. 592-6.
357. Remick, R.A., P. Jewesson, and R.W. Ford, *Monoamine Oxidase Inhibitors in General Anesthesia: A Reevaluation.* Convulsive Therapy, 1987. **3**(3): p. 196-203.
358. Ebrahim, Z.Y., et al., *Monoamine oxidase inhibitors and elective surgery.* Cleve Clin J Med, 1993. **60**(2): p. 129-30.
359. Cameron, A.G., *Monoamine oxidase inhibitors and general anaesthesia.* Anaesthesia and Intensive Care, 1986. **14**(2): p. 210.
360. Noorily, S.H., C.B. Hantler, and E.Y. Sako, *Monoamine oxidase inhibitors and cardiac anesthesia revisited.* Southern Medical Journal, 1997. **90**(8): p. 836-8.
361. van Haelst, I.M., et al., *Antidepressive treatment with monoamine oxidase inhibitors and the occurrence of intraoperative hemodynamic events: a retrospective observational cohort study.* J Clin Psychiatry, 2012. **73**(8): p. 1103-1109.
362. Krings-Ernst, I., S. Ulrich, and M. Adli, *Antidepressant treatment with MAO-inhibitors during general and regional anesthesia: a review and case report of spinal anesthesia for lower extremity surgery without discontinuation of tranlycypromine.* Int J Clin Pharmacol Ther, 2013. **51**(10): p. 763-70.
363. Wells, D.G. and A.R. Bjorksten, *Monoamine oxidase inhibitors revisited.* Canadian Journal of Anaesthesia, 1989. **36**(1): p. 64-74.
364. Ghafoor, R. and F. Rasool, *Antidepressants and antipsychotics: anaesthetic implications.* Anaesthesia & Intensive Care Medicine, 2017. **18**(7): p. 340-343.
365. Bazire, S.R., *Sudden death associated with switching monoamine oxidase inhibitors.* Drug Intell Clin Pharm, 1986. **20**(12): p. 954-6.
366. Torre, L.E., R. Menon, and B.M. Power, *Prolonged serotonin toxicity with proserotonergic drugs in the intensive care unit.* Crit Care Resusc, 2009. **11**(4): p. 272-5.
367. Mattes, J.A., *Stroke resulting from a rapid switch from phenelzine to tranlycypromine.* J Clin Psychiatry, 1998. **59**(7): p. 382.
368. Safferman, A.Z. and S.J. Masiar, *Central nervous system toxicity after abrupt Monoamine on today's inhibitor switch: a case report.* Pharmacotherapy, 1992. **26**: p. 337-338.
369. Schrire, I., *Collapse after Parstelin.* Brit Med J, 1963. **Collapse after Parstelin**(ii): p. 748.
370. Chandler, J.D., *Switching MAOIs.* J Clin Psychopharmacol, 1987. **7**: p. 438.

Check web site for latest V

64

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

371. Jefferson, J.W., *Problems with switching rapidly from one MAOI to another*. J Clin Psychiatry, 1998. **59**(2): p. 87.
372. True, B.L., B. Alexander, and B. Carter, *Switching monoamine oxidase inhibitors*. Drug Intell Clin Pharm, 1985. **19**(11): p. 825-7.
373. True, B.L., B. Alexander, and B.L. Carter, *Comment: Switching MAO inhibitors*. Drug Intell Clin Pharm, 1986. **20**(5): p. 384-5.
374. Gelenberg, A., *Switching MAOIs*. Biological Therapies in Psychiatry, 1984. **7**(9): p. 33-36.
375. Gelenberg, A., *Switching MAOIs---the sequel*. Biological Therapies in Psychiatry, 1985. **8**: p. 41.
376. Szuba, M.P., M. Hornig-Rohan, and J.D. Amsterdam, *Rapid conversion from one monoamine oxidase inhibitor to another [see comments]*. Journal of Clinical Psychiatry, 1997. **58**(7): p. 307-10.
377. Brodie, B.B., et al., *Interaction between desipramine, tyramine, and amphetamine at adrenergic neurones*. Br J Pharmacol, 1968. **34**(3): p. 648-58.
378. Matsumoto, C., E. Costa, and B.B. Brodie, *The interaction of tyramine and desmethylimipramine (DMI) with NE stores of rat hearts*. Pharmacologist, 1964. **6**: p. 206.
379. Freyschuss, U., F. Sjoqvist, and D. Tuck, *Tyramine pressor effects in man before and during treatment with nortriptyline or ECT: Correlation between plasma level and effect of nortriptyline*. European Journal of Clinical Pharmacology, 1970. **2**(33): p. 72-78.
380. Kline, N.S., et al., *Protection of patients on MAOIs against hypertensive crises*. J Clin Psychopharmacol, 1981. **1**(6): p. 410-1.
381. Pare, C.M., et al., *Will amitriptyline prevent the "cheese" reaction of monoamine-oxidase inhibitors?* Lancet, 1982. **2**(8291): p. 183-6.
382. Pare, C.M., et al., *Attempts to attenuate the 'cheese effect'. Combined drug therapy in depressive illness*. Journal of Affective Disorders, 1985. **9**(2): p. 137-41.
383. Chalon, S.A., et al., *Duloxetine increases serotonin and norepinephrine availability in healthy subjects: a double-blind, controlled study*. Neuropsychopharmacology, 2003. **28**(9): p. 1685-93.
384. Rudnick, G., *Mechanisms of biogenic amine transporters*, in *Neurotransmitter Transporters: Structure, Function and Regulation*, M.E.A. Reith, Editor. 1997: Humana Press, Totowa, NJ. p. 73– 100.
385. Bevan, P., et al., *Comparison of the responses of single cortical neurones to tyramine and noradrenaline: effects of desipramine*. Br J Pharmacol, 1978. **63**(4): p. 651-7.
386. Ghose, K., et al., *Studies of the interaction of desmethylimipramine with tyramine in man after a single oral dose, and its correlation with plasma concentration*. Br J Clin Pharmacol, 1976. **3**(2): p. 334-7.

Check web site for latest V

65

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

387. Reimann, I.W., et al., *Oxaprotiline: enantioselective noradrenaline uptake inhibition indicated by intravenous amine pressor tests but not alpha 2-adrenoceptor binding to intact platelets in man*. Eur J Clin Pharmacol, 1993. **44**(1): p. 93-5.
388. Graefe, K.H., et al., *Sympathomimetic effects of MIBG: comparison with tyramine*. J Nucl Med, 1999. **40**(8): p. 1342-51.
389. Dostert, P., et al., *Reboxetine prevents the tranylcypromine-induced increase in tyramine levels in rat heart*. Journal of Neural Transmission, 1994. **41**: p. 149-53.
390. Doggrell, S.A. and G.N. Woodruff, *Effects of antidepressant drugs on noradrenaline accumulation and contractile responses in the rat anococcygeus muscle*. British Journal of Pharmacology, 1977. **59**(3): p. 403-9.
391. Burkard, W., et al., *Interaction of moclobemide and tricyclic antidepressants with the tyramine pressor effect in rats*. Psychopharmacology (Berl), 1992. **106 Suppl**: p. S35-6.
392. Oates, J.A., et al., *The relative efficacy of guanethidine, methyl dopa and pargyline as antihypertensive agents*. N Engl J Med, 1965. **273**(14): p. 729-34.
393. Van Dyne, J.R., *Pargyline Hydrochloride in Treatment of Resistant Hypertension*. N Y State J Med, 1965. **65**: p. 1672-5.
394. Colliard, M., A. Michelet, and P. Tcherdakoff, *[Treatment of certain refractory arterial hypertension with a monoamine oxidase inhibitor]*. Archives des Maladies du Coeur et des Vaisseaux, 1981. **74 Spec No**: p. 99-106.
395. Lavin, M.R., A. Mendelowitz, and M.H. Kronig, *Spontaneous hypertensive reactions with monoamine oxidase inhibitors*. Biol Psychiatry, 1993. **34**(3): p. 146-51.
396. Fallon, B., et al., *'Spontaneous' hypertensive episodes with monoamine oxidase inhibitors*. J Clin Psychiatry, 1988. **49**(4): p. 163-5.
397. Keck, P.E., Jr., et al., *Acute cardiovascular response to monoamine oxidase inhibitors: a prospective assessment*. J Clin Psychopharmacol, 1989. **9**(3): p. 203-6.
398. Plass, H.F., *Monoamine-Oxidase Inhibitor Reactions Simulating Pheochromocytoma Attacks*. Annals of Internal Medicine, 1964. **61**: p. 924-7.
399. Bosscher, M.R., et al., *An adrenal mass and increased catecholamines: monoamine oxidase or pheochromocytoma effect?* J Clin Med Res, 2015. **7**(3): p. 199-201.
400. Cook, R.F. and D. Katritsis, *Hypertensive crisis precipitated by a monoamine oxidase inhibitor in a patient with phaeochromocytoma*. BMJ, 1990. **300**(6724): p. 614.
401. De Villiers, J.C., *Intracranial haemorrhage in patients treated with monoamine oxidase inhibitors*. British Journal of Psychiatry, 1966. **112**(483): p. 109-18.
402. Patel, K.K., et al., *Characteristics and Outcomes of Patients Presenting With Hypertensive Urgency in the Office Setting*. JAMA Intern Med, 2016. **176**(7): p. 981-8.
403. Palatini, P., et al., *Blood pressure changes during heavy-resistance exercise*. J Hypertens Suppl, 1989. **7**(6): p. S72-3.

Check web site for latest V

66

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

404. Oh, D.-J., H.-O. Hong, and B.-A. Lee, *The effects of strenuous exercises on resting heart rate, blood pressure, and maximal oxygen uptake*. Journal of exercise rehabilitation, 2016. **12**(1): p. 42.
405. de Sousa, N.M., et al., *Continuous blood pressure response at different intensities in leg press exercise*. European journal of preventive cardiology, 2014. **21**(11): p. 1324-1331.
406. MacDougall, J.D., et al., *Arterial blood pressure response to heavy resistance exercise*. J Appl Physiol, 1985. **58**(3): p. 785-90.
407. Haykowsky, M.J., J.M. Findlay, and A.P. Ignaszewski, *Aneurysmal subarachnoid hemorrhage associated with weight training: three case reports*. Clin J Sport Med, 1996. **6**(1): p. 52-5.
408. Ahsberg, K., et al., *Hospitalisation of and mortality from bleeding peptic ulcer in Sweden: a nationwide time-trend analysis*. Aliment Pharmacol Ther, 2011. **33**(5): p. 578-84.
409. Jiang, H.Y., et al., *Use of selective serotonin reuptake inhibitors and risk of upper gastrointestinal bleeding: a systematic review and meta-analysis*. Clin Gastroenterol Hepatol, 2015. **13**(1): p. 42-50 e3.
410. Anglin, R., et al., *Risk of upper gastrointestinal bleeding with selective serotonin reuptake inhibitors with or without concurrent nonsteroidal anti-inflammatory use: a systematic review and meta-analysis*. Am J Gastroenterol, 2014. **109**(6): p. 811-9.
411. Andrade, C., et al., *Serotonin reuptake inhibitor antidepressants and abnormal bleeding: a review for clinicians and a reconsideration of mechanisms*. J Clin Psychiatry, 2010. **71**(12): p. 1565-75.
412. Carvajal, A., et al., *Selective serotonin reuptake inhibitors and gastrointestinal bleeding: a case-control study*. PLoS One, 2011. **6**(5): p. e19819.
413. Richter, J.A., et al., *Bleeding after percutaneous endoscopic gastrostomy is linked to serotonin reuptake inhibitors, not aspirin or clopidogrel*. Gastrointest Endosc, 2011. **74**(1): p. 22-34 e1.
414. Dall, M., et al., *Re-prescribing of causative drugs in persons discharged after serious drug-induced upper gastrointestinal bleeding*. Aliment Pharmacol Ther, 2012. **35**(8): p. 948-54.
415. Lee, Y.C., et al., *Antidepressant use and the risk of upper gastrointestinal bleeding in psychiatric patients: a nationwide cohort study in Taiwan*. J Clin Psychopharmacol, 2012. **32**(4): p. 518-24.
416. Marik, P.E. and R. Rivera, *Hypertensive emergencies: an update*. Curr Opin Crit Care, 2011. **17**(6): p. 569-80.
417. Flanigan, J.S. and D. Vitberg, *Hypertensive emergency and severe hypertension: what to treat, who to treat, and how to treat*. Med Clin North Am, 2006. **90**(3): p. 439-51.
418. Migneco, A., et al., *Hypertensive crises: diagnosis and management in the emergency room*. Eur Rev Med Pharmacol Sci, 2004. **8**(4): p. 143-52.
419. Feldstein, C., *Management of hypertensive crises*. Am J Ther, 2007. **14**(2): p. 135-9.

Check web site for latest V

67

[Consider a donation to maintain the PsychoTropical website](#)

Address for correspondence. Dr Ken Gillman: ken.psychotropic@gmail.com

420. Marik, P.E. and J. Varon, *Hypertensive crises: challenges and management*. Chest, 2007. **131**(6): p. 1949-62.
421. Schwartz, M., et al., *Oral nifedipine in the treatment of hypertensive urgency: cerebrovascular accident following a single dose*. Arch Intern Med, 1990. **150**(3): p. 686-7.
422. Bulling, M. and R. Burns, *Occipital cortical "angina" induced by nifedipine*. Med J Aust, 1988. **148**(5): p. 266.
423. Burton, T.J. and I.B. Wilkinson, *The dangers of immediate-release nifedipine in the emergency treatment of hypertension*. J Hum Hypertens, 2008. **22**(4): p. 301-2.
424. Grossman, E., et al., *Should a moratorium be placed on sublingual nifedipine capsules given for hypertensive emergencies and pseudoemergencies?* Jama, 1996. **276**(16): p. 1328-1331.
425. McCormack, D. and N. Buckley, *Psychostimulant poisoning*. Australian Prescriber, 2006. **29**: p. 109–11.
426. Murray, L., et al., *Toxicology handbook*. 2011: Elsevier Australia.
427. Grossman, E., et al., *Antianxiety treatment in patients with excessive hypertension*. Am J Hypertens, 2005. **18**(9 Pt 1): p. 1174-7.
428. Yilmaz, S., et al., *Comparison of alprazolam versus captopril in high blood pressure: a randomized controlled trial*. Blood Press, 2011. **20**(4): p. 239-43.